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PFOS: A FROG EMBRYO TERATOGENESIS ASSAY – XENOPUS (FETAX)

FINAL REPORT

WILDLIFE INTERNATIONAL, LTD. PROJECT NUMBER: 454A-116

3M LAB REQUEST NO. U2723

ASTM Method E 1439-91

AUTHORS:

Susan J. Palmer Henry O. Krueger, Ph.D.

STUDY INITIATION DATE: May 11, 2000

STUDY COMPLETION DATE: April 6, 2001

SUBMITTED TO:

3M Corporation Environmental Laboratory 935 Bush Avenue St. Paul, Minnesota 55144

Wildlife International, Ltd.

8598 Commerce Drive Easton, Maryland 21601 (410) 822-8600

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GOOD LABORATORY PRACTICE COMPLIANCE STATEMENT

SPONSOR:

3M Corporation

TITLE:

PFOS: A Frog Embryo Teratogenesis Assay - Xenopus (FETAX)

WILDLIFE INTERNATIONAL, LTD. PROJECT NUMBER: 454A-116

STUDY COMPLETION: April 6, 2001

This study was not conducted in compliance with Good Laboratory Practice Standards as published by the U.S. Environmental Protection Agency in 40 CFR Parts 160 and 792, 17 August 1989. Data collection, record keeping, training and facility records at Wildlife International, Ltd. complied with Good Laboratory Practices, with the following exceptions:

The test substance was not characterized in accordance with full GLP compliance prior to the study; however, the characterization was performed according to 3M Standard Operating Procedures and Methods, and all raw data are being maintained in the 3M archives. The test substance has been recharacterized in accordance with GLP on September 7, 2000.

The stability of the test substance under conditions of storage at the test site was not determined in accordance with Good Laboratory Practice Standards.

The phase of the study conducted by University of Maryland was not conducted in compliance with Good Laboratory Practices.

STUDY DIRECTOR:

Susan J. Palmer

Senior Biologist

DATE

4-6-01

SPONSOR APPROVAL:

DATE

-3-

QUALITY ASSURANCE STATEMENT

This study was examined for compliance with Good Laboratory Practice Standards as published by the U.S. Environmental Protection Agency, 40 CFR Parts 160 and 792, 17 August 1989. The dates of all inspections and audits and the dates that any findings were reported to the Study Director and Laboratory Management were as follows:

ACTIVITY:	DATE CONDUCTED:	DATE REPO STUDY DIRECTOR:	RTED TO: MANAGEMENT:
Test Substance Preparation	May 12, 2000	May 12, 2000	May 18, 2000
Analytical Data and Draft Report	July 12 & 13, 2000	July 13, 2000	July 14, 2000

Timothy A. Springer, Ph.D.

Manager, Regulatory and Technical Support

4/6/01 DATE -4-

REPORT APPROVAL

SPONSOR:

3M Corporation

TITLE:

PFOS: A Frog Embryo Teratogenesis Assay - Xenopus (FETAX)

WILDLIFE INTERNATIONAL, LTD. PROJECT NUMBER: 454A-116

STUDY DIRECTOR:

Susan J. Palmer

Senior Biologist

DATE

MANAGEMENT:

Henry O. Krueger, Ph.D.

Director, Aquatic Toxicology and

Non-Target Plants

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SUMMARY

SPONSOR: 3M Corporation SPONSOR'S REPRESENTATIVE: Rochelle R. Robideau LOCATION OF STUDY, RAW DATA AND A COPY OF THE Wildlife International, Ltd. FINAL REPORT: Easton, Maryland 21601 WILDLIFE INTERNATIONAL, LTD. PROJECT NUMBER: 454A-116 PFOS: A Frog Embryo Teratogenesis Assay - Xenopus (FETAX) STUDY: PFOS (Perfluorooctanesulfonate, Potassium Salt) TEST SUBSTANCE: IUPAC Name: 1-Octanesulfonic acid, 1, 1, 2, 2, 3, 3, 4, 4, 5, 5, 6, 6, 7,7,8,8,8-heptadecafluoro-potassium salt; CAS #2795-39-3 6-aminonicotinamide REFERENCE SUBSTANCE: Negative Control, 1.82, 3.07, 5.19, 8.64, 14.4 and 24.0 mg a.i./L NOMINAL TEST CONCENTRATIONS: and 5.5 and 2500 mg/L 6-aminonicotinamide 1st Assay: Negative Control, 2.00, 2.83, 4.73, 7.90, 14.7 and MEAN MEASURED TEST 24.6 mg a.i./L CONCENTRATIONS: 2nd Assay: Negative Control, 1.91, 3.04, 4.82, 7.97, 13.3 and 23.1 mg a.i./L 3rd Assay: Negative Control, 1.93, 3.27, 5.25, 8.26, 14.0 and 23.9 mg a.i./L 1st Assay: Experimental Start - May 15, 2000 TEST DATES: Biological Termination - May 19, 2000 Experimental Termination - May 22, 2000 2nd and 3rd Assays: Experimental Start - May 22, 2000 Biological Termination - May 26, 2000 Experimental Termination - May 30, 2000 96 Hours LENGTH OF EXPOSURE:

TEST ORGANISM:	South African Clawed Frog (Xenopus laevis)	
SOURCE OF TEST ORGANISMS:	University of Maryland Wye Research and Education Center Queenstown, Maryland 21658	
AGE OF TEST ORGANISMS:	Embryos	

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SUMMARY

- Continued -

RESULTS (BASED ON MEAN MEASURED CONCENTRATIONS):	1st Assay:	2 nd Assay:	3rd Assay:
96-HOUR LC ₅₀ :	13.8 mg a.i./L	17.6 mg a.i./L	15.3 mg a.i./L
95% CONFIDENCE LIMITS:	12.38 – 15.31 mg a.i./L	15.53 – 19.96 mg a.i./L	13.13 – 17.75 mg a.i./L
96-HOUR EC50:	12.1 mg a.i./L	17.6 mg a.i./L	16.8 mg a.i./L
95% CONFIDENCE LIMITS:	10.00 - 14.58 mg a.i./L	13.49 – 22.88 mg a.i./L	12.35 – 22.82 mg a.i./L
MINIMUM CONCENTRATION TO INHIBIT GROWTH (MCIG):	Not calculable	7.97 mg a.i./L	8.26 mg a.i./L
TERATOGENIC INDEX (TI):	1.1	1.0	0.9

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INTRODUCTION

This study was conducted in three assays by Wildlife International, Ltd. for 3M Corporation at the University of Maryland Wye Research and Education Center (UM-WREC) in Queenstown, Maryland and at the Wildlife International, Ltd. aquatic toxicology facility in Easton, Maryland. The in-life phase of the first assay was conducted from May 15 to May 19, 2000, while the second and third assays were concurrently conducted from May 22 to May 26, 2000. Raw data generated by Wildlife International, Ltd., data obtained from the University of Maryland, and a copy of the final report are filed under Project Number 454A-116 in archives located on the Wildlife International, Ltd. site.

OBJECTIVE

The objective of this study was to determine the possible developmental toxicity of <u>PFOS</u> (Perfluorooctanesulfonate, Potassium Salt), also referred to as test substance U2723, on the embryos of the South African clawed frog (*Xenopus laevis*) during a 96-hour exposure period.

EXPERIMENTAL DESIGN

Three definitive assays were conducted with PFOS using the frog embryo teratogenesis assay – *Xenopus* (FETAX). During each assay, *Xenopus* embryos were exposed to a geometric series of six test concentrations (PFOS in FETAX solution), a negative control (FETAX solution only), and two concentrations of a reference toxicant (6-aminonicotinamide in FETAX solution) for 96 hours. Two replicate test chambers were maintained in each treatment group and four replicate test chambers were maintained in each control group. Each test chamber contained 25 cmbryos for a total of 50 embryos per treatment group and 100 embryos per control group. An abiotic replicate at the highest concentration was included in each assay for analytical sampling at 96 hours.

Nominal test concentrations were selected in consultation with the Sponsor, and were based upon the results of an exploratory range finding toxicity test. Nominal test concentrations selected were 1.82, 3.07, 5.19, 8.64, 14.4 and 24.0 mg active ingredient (a.i.)/L. Mean measured test concentrations were determined at Wildlife International, Ltd. from samples of test solution collected from each treatment and the negative control group prior to and at the end of each assay.

The test solutions were prepared at Wildlife International, Ltd. and were delivered to UM-WREC prior to each assay. At UM-WREC, the embryos were randomly assigned to test chambers containing the appropriate FETAX solution and were observed for 96 hours. The FETAX solutions in the test chambers were renewed every 24 hours, and mortality observations and general water quality measurements were made at each 24-hour renewal period. At the end of the 96-hour exposure period, each surviving embryo was examined for malformations using a dissecting microscope. An LC50 value for mortality and an EC50 value for malformation were calculated at 96 hours for each assay. When possible, the minimum concentration to inhibit growth (MCIG) and the teratogenic index (TI) were calculated for each assay.

MATERIALS AND METHODS

The study was conducted based on the procedures outlined in the protocol, "PFOS: A Frog Embryo Teratogenesis Assay – Xenopus (FETAX)". The protocol was based on procedures outlined in ASTM Standard E1439-91 Standard Guide for Conducting the Frog Embryo Teratogenesis Assay – Xenopus (FETAX) (1) and ASTM Standard E729-88a Standard Guide for Conducting Acute Toxicity Tests with Fishes, Macroinvertebrates and Amphibians (2).

Test and Reference Substances

The test substance was received from 3M Corporation on October 29, 1998 and was assigned Wildlife International, Ltd. identification number 4675. The test substance was described as a white powder. It was identified as FC-95 from lot number 217 (T-6295). Information provided by the Sponsor indicated a purity of 98.9%, and an expiration date of 2008. The test substance was reanalyzed by the Sponsor and the Certificate of Analysis dated September 7, 2000 indicated a purity of 86.9% and an expiration date of August 31, 2001. The test substance was stored at ambient room temperature.

The reference substance was maintained by UM-WREC. It was identified as 6-aminonicotinamide (CAS #329-89-5; Sigma; Lot No. 125H0006).

Preparation of Test Concentrations

Prior to each assay, FETAX solution was prepared at UM-WREC and delivered to Wildlife International, Ltd. for the preparation of the stock and test solutions. The second and third assays were conducted concurrently using common test solutions. A primary stock solution was prepared in FETAX solution at a concentration of 48 mg a.i./L. The primary stock solution was mixed by sonication and stirring to aid in the solubilization of the

test substance. The primary stock was proportionally diluted with FETAX solution to prepare the six test concentrations. Nominal test concentrations were 1.82, 3.07, 5.19, 8.64, 14.4 and 24.0 mg a.i./L, based on a test substance purity of 86.9%. The test solutions were delivered to UM-WREC prior to the start of each assay. The reference toxicant solutions were prepared at UM-WREC.

Analytical Chemistry

Samples were collected from each test solution and the negative control solution at preparation, and at the termination of each assay (96 hours) to measure concentrations of the test substance. Samples of test solution from the abiotic replicates also were collected at 96 hours. The samples were analyzed at Wildlife International, Ltd. Analytical procedures used in the analysis of the samples are provided in Appendix II.

In-Life Phase of Assays

Details of the in-life phase of the three assays conducted at UM-WREC are included in Appendix I.

RESULTS AND DISCUSSION

Measurement of Test Concentrations

Results of analyses to measure concentrations of PFOS in FETAX solution samples collected during the three assays are presented in Table 1 and in the analytical chemistry report (Appendix II). Nominal concentrations selected for use in this study were 1.82, 3.07, 5.19, 8.64, 14.4 and 24.0 mg a.i./L. In the first assay, the measured concentrations of PFOS in test solution samples collected prior to initiation of exposure of the test organisms ranged from 112 to 141% of the nominal concentrations. Samples collected at test termination had a measured concentration range of 54.7 to 98.6% of nominal values. The sample from the abiotic 24.0 mg a.i./L treatment group was comparable to samples from the 24.0 mg a.i./L treatment group with the frog embryos present.

For the second and third assays, common test solutions were prepared. The measured concentrations of PFOS in test solution samples collected prior to initiation of exposure of the test organisms ranged from 95.8 to 117% of the nominal concentrations. In the second assay, samples collected at test termination had a measured concentration range of 80.7 to 112% of nominal values. The sample from the abiotic 24.0 mg a.i./L treatment group was comparable to samples from the 24.0 mg a.i./L treatment group with the frog embryos present. In the third assay, samples collected at test termination had a measured concentration range of 93.4 to 114% of nominal values. The sample from the abiotic 24.0 mg a.i./L treatment group was comparable to samples from the 24.0 mg a.i./L treatment group was comparable to samples from the 24.0 mg a.i./L treatment group with the frog embryos present.

When measured concentrations of the samples analyzed prior to and at the end of each assay were averaged, the mean measured concentrations for the first assay were 2.00, 2.83, 4.73, 7.90, 14.7 and 24.6 mg a.i./L, representing 110, 92.2, 91.1, 91.4, 102 and 103% of nominal concentrations, respectively. The mean measured concentrations for the second assay were 1.91, 3.04, 4.82, 7.97, 13.3 and 23.1 mg a.i./L, representing 105, 99.0, 92.9, 92.2, 92.4 and 96.3% of nominal concentrations, respectively. The mean measured concentrations for the third assay were 1.93, 3.27, 5.25, 8.26, 14.0 and 23.9 mg a.i./L, representing 106, 107, 101, 95.6, 97.2 and 99.6% of nominal concentrations, respectively. The results of the study were based on the mean measured test concentrations for each assay.

Definitive Assay Results

The results of the three definitive FETAX assays conducted with PFOS at the UM-WREC are included in Appendix I and are summarized in Table 2. Exposure to PFOS caused significant embryo mortality at the two highest test concentrations (14.4 and 24.0 mg a.i./L nominal concentration) in each of the three assays. Most of the embryo mortality appeared to be caused by the gut coiling through the body wall at the two highest concentrations. Embryo survival was ≥90% at all test concentrations up to 8.64 mg a.i./L nominal concentration. The 96-hour LC50 values for PFOS ranged from 13.8 to 17.6 mg a.i./L mean measured concentration in the three assays (Table 2).

There was a positive correlation between PFOS exposure and malformations in each of the three assays, with increasing concentrations causing an increase in the incidence of malformed embryos. The 96-hour EC50 values, based on the incidence of malformed embryos, ranged from 12.1 to 17.6 mg a.i./L mean measured concentration in the three assays (Table 2). The most common types of malformations caused by exposure to PFOS were improper gut coiling, edema, notochord abnormalities and facial abnormalities.

Growth of the embryos was not effected by exposure to PFOS in the first assay. In the second and third assays, the minimum concentration inhibiting growth (MCIG) was 8.64 mg a.i./L nominal concentration. This reduction in growth may be attributed more to the increased incidence of malformed embryos in the test concentrations than to an overall decrease in growth. The teratogenic index (TI), defined as the 96-hour LC50 value for mortality divided by the 96-hour EC50 value for malformed embryos, provides an estimate of the teratogenic risk associated with PFOS. The TI's in the three assays ranged from 0.9 to 1.1, indicating the PFOS has a low potential to be a developmental hazard.

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CONCLUSIONS

The 96-hour LC50 value for mortality for *Xenopus laevis* exposed to PFOS in each of three assays was 13.8, 17.6 and 15.3 mg a.i./L mean measured concentration, respectively. The 96-hour EC50 value for embryo malformation in each of three assays was 12.1, 17.6 and 16.8 mg a.i./L mean measured concentration, respectively. The MCIG, determined in the second and third assays, was 7.97 and 8.26 mg a.i./L mean measured concentration, respectively. The TI's ranged from 0.9 to 1.1 in the three assays, indicating the PFOS has a low potential to be a developmental hazard.

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REFERENCES

- 1 ASTM Standard E1439-91. 1998. Standard Guide for Conducting the Frog Embryo Teratogenesis Assay Xenopus (FETAX). American Society for Testing and Materials. Philadelphia, Pennsylvania.
- 2 ASTM Standard E729-88a. 1994. Standard Guide for Conducting Acute Toxicity Tests with Fishes, Macroinvertebrates, and Amphibians. American Society for Testing and Materials. Philadelphia, Pennsylvania.

Table 1 Summary of Analytical Chemistry Data

Nominal Test	Sampling		1st Assay			2 nd Assay ^I			3 rd Assay ^I	
Concentration (mg a.i./L)	Time (Day)	Measured Concentration (mg a.i./L)	Mean Measured Concentration (mg a.i./L)	Percent of Nominal	Measured Concentration (mg a.i./L)	Mean Measured Concentration (mg a.i./L)	Percent of Nominal	Measured Concentration (mg a.i./L)	Mean Measured Concentration (mg a.i./L)	Percent of Nominal
Negative Control	0 ² 4 ³	<loq<sup>4 <loq< td=""><td><loq< td=""><td></td><td><too <too< td=""><td><loq< td=""><td></td><td><loq <loq< td=""><td><loq< td=""><td></td></loq<></td></loq<></loq </td></loq<></td></too<></too </td></loq<></td></loq<></loq<sup>	<loq< td=""><td></td><td><too <too< td=""><td><loq< td=""><td></td><td><loq <loq< td=""><td><loq< td=""><td></td></loq<></td></loq<></loq </td></loq<></td></too<></too </td></loq<>		<too <too< td=""><td><loq< td=""><td></td><td><loq <loq< td=""><td><loq< td=""><td></td></loq<></td></loq<></loq </td></loq<></td></too<></too 	<loq< td=""><td></td><td><loq <loq< td=""><td><loq< td=""><td></td></loq<></td></loq<></loq </td></loq<>		<loq <loq< td=""><td><loq< td=""><td></td></loq<></td></loq<></loq 	<loq< td=""><td></td></loq<>	
1.82	0 4	2.58 1.42	2.00	110	1.77 2.04	1.91	105	1.77 2.08	1.93	106
3.07	0 4	3.94 1.72	2.83	92.2	3.59 2.49	3.04	99.0	3.59 2.94	3.27	107
5.19	0 4	6.62 2.84	4.73	91.1	5.45 4.18	4.82	92.9	5.45 5.05	5.25	101
8.64	0 4	10.7 5.09	7.90	91.4	8.43 7.51	7.97	92.2	8.43 8.09	8.26	95.6
14.4	0 4	18.5 10.8	14.7	102	14.5 12.1	13.3	92.4	14.5 13.5	14.0	97.2
24.0	0 4	26.9 22.3	24.6	103	23.0 23.1	23.1	96.3	23.0 24.7	23.9	99.6
24.0 (abiotic)	4	23.7	-	98.6	23.9	-	99.6	24.1	-	100

The 2nd and 3rd assays were conducted concurrently using common batches of test solution prepared for Day 0.

Day 0 samples were collected from the test solutions at preparation prior to transfer to UM-WREC.

Day 4 samples were collected from the test solutions at the termination of each assay at UM-WREC and transferred to Wildlife International, Ltd. for analysis.

⁴ The limit of quantitation (LOQ) was 0.240 mg a.i./L.

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Table 2 LC50, EC50, MCIG and TI Values 1

Parameter	1st Assay	2 nd Assay	3 rd Assay
96-Hour LC50 (mg a.i./L):	13.8	17.6	15.3
95% Confidence Limits (mg a.i./L):	12.38 – 15.31	15.53 – 19.96	13.13 – 17.75
96-Hour EC50 (mg a.i./L):	12.1	17.6	16.8
95% Confidence Limits (mg a.i./L):	10.00 - 14.58	13.49 – 22.88	12.35 - 22.82
Minimum Concentration Inhibiting Growth (MCIG) (mg a.i./L):	2	7.97	8.26
Teratogenic Index (TI):	1.1	1.0	0.9

Values are based on the mean measured concentrations for each assay.
 The MCIG could not be calculated because PFOS had no effect on embryonic growth at any of the test concentrations with surviving embryos in the 1st assay.

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APPENDIX I

Certificate of Analysis

INTERIM CERTIFICATE OF ANALYSIS

Revision 1(9/7/00)

Centre Analytical Laboratories COA Reference #: 023-018A

3M Product: PFOS, Lot 217 Reference #: SD-018 Purity: 86.9%

Test Name	Purity: 86.9% Specifications	Result	
Purity ¹	NATION OF BUILDING	86.9%	
Appearance	White Crystalline Powder	Conforms	
Identification NMR		Positive	
Metals (ICP/MS) 1. Calcium 2. Magnesium 3. Sodium 4. Potassium ² 5. Nickel 6. Iron 7. Manganese		1. 0.005 wt/wt.% 2. 0.001 wt/wt.% 3. 1.439 wt/wt.% 4. 6.849 wt/wt.% 5. <0.001 wt/wt.% 6. 0.005 wt/wt.% 7. <0.001 wt/wt.%	
Total % Impurity (NMR)	· 在刘克里等的特别是2.5000000000000000000000000000000000000	1.93 wt./wt.%	
Total % Impurity (LC/MS)		8.41 wt/wt.%	
Total % Impurity (GC/MS)	Berger Land	None Detected	
Related Compounds – POAA		0.33 wt./wt.%	
Residual Solvents (TGA)		None Detected	
Purity by DSC		Not Applicable ³	
Inorganic Anions (IC) 1. Chloride 2. Fluoride 3. Bromide 4. Nitrate 5. Nitrite 6. Phosphate 7. Sulfate ⁴		1. <0.015 wt./wt.% 2. 0.59 wt./wt.% 3. <0.040 wt./wt.% 4. <0.009 wt./wt.% 5. <0.006 wt./wt.% 6. <0.007 wt./wt.% 7. 8.76 wt./wt.%	
Organic Acids ⁵ (IC) 1. TFA 2. PFPA 3. HFBA 4. NFPA		1. <0.1 wt/wt.% 2. <0.1 wt/wt.% 3. 0.10 wt/wt.% 4. 0.28 wt/wt.%	
Elemental Analysis ⁶ : 1. Carbon 2. Hydrogen 3. Nitrogen 4. Sulfur 5. Fluorine	1. Theoretical Value = 17.8% 2. Theoretical Value = 0% 3. Theoretical Value = 0% 4. Theoretical Value = 5.95% 5. Theoretical Value = 60%	1. 12.48 wt./wt.% 2. 0.244 wt./wt.% 3. 1.74 wt./wt.% 4. 8.84 wt./wt.% 5. 54.1 wt./wt.%	

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INTERIM CERTIFICATE OF ANALYSIS Centre Analytical Laboratories COA Reference #: 023-018A

Date of Last Analysis: 08/31/00

Expiration Date: 08/31/01

Storage Conditions: Frozen ≤-10°C

Re-assessment Date: 08/31/01

¹Purity = 100% - (sum of metal impurities, 1.45% +LC/MS impurities, 8.41%+Inorganic Fluoride, 0.59%+NMR impurities, 1.93%+organic acid impurities, 0.38%+POAA, 0.33%)

Total impurity from all tests = 13.09% Purity = 100% - 13.09% = 86.9%

Trifluoroacetic acid
HFBA Heptafluorobutyric acid
NFPA Nonofluoropentanoic acid
PFPA Pentafluoropropanoic acid

⁶Theoretical value calculations based on the empirical formula, C₈F₁₇SO₃K⁺(MW=538)

This work was conducted under EPA Good Laboratory Practice Standards (40 CFR 160).

²Potassium is expected in this salt form and is therefore not considered an impurity.

³Purity by DSC is generally not applicable to materials of low purity. No endotherm was observed for this sample.

⁴Sulfur in the sample appears to be converted to SO₄ and hence detected using the inorganic anion method conditions. The anion result agrees well with the sulfur determination in the elemental analysis, lending confidence to this interpretation. Based on the results, the SO₄ is not considered an impurity.

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INTERIM CERTIFICATE OF ANALYSIS Centre Analytical Laboratories COA Reference #: 023-018A

LC/MS Purity Profile:

Impurity	wt./wt. %
C4	1.22
C5	1.33
C6	4.72
C7	1.14
Total	8.41

Note: The C4 and C6 values were calculated using the C4 and C6 standard calibration curves, respectively. The C5 value was calculated using the average response factors from the C4 and C6 standard curves. Likewise, the C7 value was calculated using the average response factors from the C6 and C8 standard curves.

Prepared By:		
	David S. Bell	Date
	Scientist, Centre Analytical Laboratories	
Reviewed By:	Secretary and the secretary an	244
€₹2	John Flaherty	Date
	Laboratory Manager, Centre Analytical Lal	boratories

COA023-018A

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APPENDIX II

Evaluation of the Toxicity and Teratogenicity of Chemical U2723 Using the Frog Embryo Teratogenesis Assay - Xenopus (FETAX)

Report No. WREC-00-02

EVALUATION OF THE TOXICITY AND TERATOGENICITY OF CHEMICAL U2723 USING THE FROG EMBRYO TERATOGENESIS ASSAY-XENOPUS (FETAX)

Final Report

Prepared for

Wildlife International, Ltd. 8598 Commerce Drive Easton, MD 21601

Prepared by

Steven D. Turley and Dennis T. Burton

University of Maryland Wye Research and Education Center P.O. Box 169 Queenstown, MD 21658

July 15, 2000

Revision Date:

March 19, 2001

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1. INTRODUCTION

The evaluation of the potential toxicity and teratogenicity of chemical U2723 was initiated at the request of Mr. David A. Palmer, Wildlife International, Ltd., Easton, Maryland. The study was conducted by Mr. Steven D. Turley and Dr. Dennis T. Burton of the University of Maryland Wye Research and Education Center, Queenstown, Maryland. The test program was performed in accordance with the "Revised Statement of Work for Evaluating the Potential Teratogenicity of U2723 Using 96-Hour Frog Embryo Teratogenesis Assay-Xenopus (FETAX)", submitted to and approved by Wildlife International, Ltd., March 30, 2000 (Turley, 2000). The potential toxicity and teratogenicity of chemical U2723 was evaluated by the Frog Embryo Teratogenesis Assay-Xenopus (FETAX). Three 96-h definitive FETAX assays were conducted on chemical U2723.

2. MATERIALS AND METHODS

Three definitive FETAX assays were conducted on chemical U2723 using the frog embryo teratogenesis assay-Xenopus (FETAX). The assay is a 96-h quantitative developmental assay used to screen for developmental toxicants in aquatic media. The assays were conducted using the static renewal (solutions renewed every 24 h) test protocol Designation E 1439-91 of the American Society for Testing and Materials (ASTM, 1998). Embryolethality (96-h LC50), malformations (96-h EC50), and growth retardation were used as endpoints in the assays.

Embryos were obtained from breeding colonies of *Xenopus laevis* at the University of Maryland Wye Research and Education Center (UMD/WREC). The UMD/WREC adult colonies were maintained in flow-through (~4 replacement volumes per day) circular polyethylene aquaria (0.91 m l.D. x 0.36 m high) with a water depth of 10 cm. Each aquarium contained a maximum of 10 adults. UMD/WREC non-chlorinated deep well water held at 23.5 ± 0.5 °C served as the culture medium. All frogs were fed every 5-6 d with commercial beef liver supplemented with liquid vitamins (PolyViSol™; Mead-Johnson Nutritionals, Evansville, Indiana). The colony was held under a photoperiod of 16 h light:8 h dark. Mating pairs were bred in the dark in 23.5 ± 0.5 °C UMD/WREC non-chlorinated water at ~70 d intervals by injecting 400 and 800 l.U. of human chorionic gonadotropin (HCG) in the dorsal lymph sac of the males and females, respectively. Amplexus occurred 4-6 h after injecting HCG; egg deposition occurred 9-12 h following HCG injection.

Embryos between normal stage 8 blastulae and normal stage 11 gastrulae were used to initiate the assays (Nieuwkoop and Faber, 1975). Embryos from separate breeding pairs were used for each test to check for variability in the sensitivity of individual clutches of embryos. All embryos were examined two times for viability. The embryos were de-jellied in a 2% L-cysteine solution (2 g of L-cysteine per 98 mL of FETAX solution). Once de-jellied, the embryos were rinsed and re-suspended in FETAX solution (ASTM, 1998). The embryos were assayed in covered 60 mm glass Petri dishes containing 10 mL of solution. All embryos were randomly assigned to the Petri dishes. Two replicates of 25 embryos/replicate were used for each of six assay treatments; four replicates were used for the controls as required by the test protocol (ASTM, 1991). The assays were conducted at 24 ± 0.2°C under a 12-h light; 12-h dark photoperiod (fluorescent lights; ~75 foot candles at the surface of the test medium) in a constant temperature environmental chamber. All Petri dishes were randomly placed in the environmental chamber used for the assays. Wildlife International, Ltd., personnel prepared and delivered stock solutions of each concentration at the start of each assay. The concentration of each solution was quantified at Wildlife International, Ltd. Two concentrations of the reference toxicant, 6-aminonicotinamide (CAS #329-89-5; Sigma; Lot No. 125H0006), were used at each assay period as outlined in the ASTM (1998) protocol.

Mortality observations and general water quality measurements were made at each 24-h renewal period. All dead embryos were removed at each observation period. At the end of the 96-h exposure period, each surviving embryo was examined for malformations using a dissecting microscope. The identification and interpretation of malformations in the embryos at 96 h were made via the atlas of Bantle et al. (1991). The head-tail length of each surviving embryo was determined via Sigma Scan's digitizing software (SPSS, Corte Madera, California). A random group of normal embryos and embryos with each type of malformation were preserved in 10% buffered formalin (pH 7) for photographing.

The Trimmed Spearman-Karber statistical procedure was used to determine the 96-h LC50 for mortality and 96-h EC50 for malformations. The minimum concentration to inhibit growth (MCIG) was determined by Bonferroni's T-Test. All statistical tests were performed using Toxstat (WEST and Gulley, 1994). A minimum probability level of 0.05 was used. The teratogenic index (TI) was calculated by dividing the LC50 by the EC50. The test acceptability criteria given in the ASTM (1998) test protocol was used for each definitive test.

3. RESULTS AND DISCUSSION

The data for definitive assays 1, 2, and 3 are summarized in Appendices 1,2, and 3, respectively. Copies of the raw data sheets and 6-aminonicotinamide reference toxicant data for assays 1, 2, and 3 are given in Appendices 4, 5, and 6, respectively. The nominal concentrations of 1.9, 3.2, 5.4, 9.0, 15.0 and 25.0 mg/L on the raw data sheets correspond to the nominal concentrations of 1.82, 3.07, 5.19, 8.64, 14.4 and 24.0 mg a.i./L in the report. Photographs of normal and malformed embryos of each type observed in the study are given in Attachment 1. Table 1 is summary of the endpoint parameters established in the three definitive assays.

Table 1. Summary of Data for the Three Assays with Chemical U2723°

Parameter	Assay 1	Assay 2	Assay 3 15.3 (13.13 - 17.75) 16.8 (12.35 - 22.82)	
LC50 (95% C. I.)	13.8 (12.38 - 15.31)	17.6 (15.53 - 19.96)		
EC50 (95% C. I.)	12.1 (10.00 - 14.58)	17.6 (13.49 - 22.88)		
MCIG	ь	7.97	8.26	
TI	1.1	1.0	0.9	

^a The LC50, EC50, and MCIG values are based on mean measured concentrations of chemical U2723 (mg a.i./L) for each assay.

Exposure to chemical U2723 caused significant embryo mortality at the two highest treatment concentrations (14.4 and 24.0 mg/L) in each of the three assays. Most of the embryo mortality appeared to be caused by the gut coiling through the body wall at the two highest assay treatments. Embryo survival was ≥90% at all exposure treatments up to 8.64 mg/L. The 96-h LC50s for chemical U2723 ranged from 13.8 to 17.6 mg/L (Table 1).

There was a positive correlation between chemical U2723 exposure and malformations in each of the three assays, with increasing concentrations causing an increase in the incidence of malformed embryos. The 96-h EC50s, based on the incidence of malformed embryos, ranged from 12.1 to 17.6 mg/L (Table 1). The most common types of malformations caused by exposure to chemical U2723 were: improper gut coiling,

The MCIG could not be calculated because chemical U2723 had no effect on embryonic growth at any of the test treatments with surviving embryos (Appendix A; Table A1- 4).

edema, notochord abnormalities and facial abnormalities.

Growth of the embryos was not affected by exposure to chemical U2723 in assay 1. In assays 2 and 3, the minimum concentration inhibiting growth (MCIG) was 7.97 and 8.26 mg/L, respectively. This reduction in growth may be attributed more to the increased incidence of malformed embryos in these test concentrations than an overall decrease in growth. Malformed embryos are generally smaller than normally developing embryos.

The teratogenic index (TI), which by definition is the 96-h LC50 divided by the 96-h EC50 (malformed embryos), provides an estimate of the teratogenic risk associated with a material (Dumont et al., 1983). TI values of 1.5 to 2.0 indicate that a material may be a potential teratogen. Materials with TI values >2.0 should be considered for further teratogenicity testing. The TI's in the current study ranged from 0.9 to 1.1 which indicate that chemical U2723 has a low potential to be a developmental hazard.

The three assays met all of the ASTM (1998) protocol test acceptability criteria, with one exception. The reference toxicant (6-aminonicotinamide) high concentration of 2,500 mg/L did not meet the ASTM protocol test acceptability criterion. A concentration of 2,500 mg/L should kill between 40 and 60% of the embryos in 96 h. All embryos died at 2,500 mg/L in all three reference toxicant tests. All three reference toxicant tests met the protocol test acceptability criteria for the low reference toxicant concentration of 5.5 mg/L which should cause between 40 and 60% malformed embryos after 96 h of exposure. The 5.5 mg/L reference toxicant tests produced an average of 55.6, 55.3, and 53.1% malformed embryos during assays 1, 2, and 3, respectively. The results of the reference toxicant concentrations in the three current tests are consistent and not at variance with previous experience in our laboratory. The survival of control embryos after 96 hours in our laboratory consistently meet the ASTM protocol acceptability criterion of >90% survival. In the current chemical U2723 assays, the average survival of the controls was 99, 99, and 100%, in assays 1, 2, and 3, respectively.

The chain of custody forms for delivery of the assay solutions from Wildlife International, Ltd. to the UMD/WREC were retained by Wildlife International, Ltd. Thus, copies of the chain of custody forms are not included in this report.

4. REFERENCES

- ASTM. 1998. Standard guide for conducting the frog embryo teratogenesis assay-Xenopus (FETAX). ASTM Designation E 1439-91. 1998 Annual Book of ASTM Standards Section 11 Water and Environmental Technology, Amer. Soc. Testing Materials, Philadelphia, PA.
- Bantle, J.A., J.N. Dumont, R.A. Finch, and G. Linder. 1991. Atlas of abnormalities. A guide for the performance of FETAX. Oklahoma State Univ., Stillwater, OK.
- Dumont, J., T. Schultz, M. Buchanan, and G. Kao. 1983. Frog embryo teratogenesis assay-Xenopus (FETAX) A short-term assay applicable to complex environmental mixtures. Pages 393-405 in: Waters, Sandhu, S.S., J. Lewtas, L. Claxton, N. Chernoff, and S. Nesnow, eds. Short-term bioassays in the analysis of complex environmental mixtures III, Plenum, New York, NY.
- Nieuwkoop, P.D. and J. Faber. 1975. Normal tables of *Xenopus laevis* (Daudin), 2[™] ed. North Holland, Amsterdam, Netherlands.
- Turley, S.D. 2000. Revised statement of work for evaluating the potential teratogenicity of U2723 using 96-hour frog embryo teratogenesis assay-Xenopus (FETAX). March 30, 2000. University of Maryland Wye Research and Education Center, Queenstown, MD.
- WEST (Western Ecosystem Technology, Inc.) and Gulley, D. D. 1994. Toxstat, v. 3.4. Western Ecosystem Technology, Inc., Cheyenne, WY.

APPENDIX 1

FROG EMBRYO TERATOGENESIS ASSAY-XENOPUS (FETAX) CONDUCTED WITH CHEMICAL U2723 (ASSAY 1)

Test Method:

ASTM Designation E 1439-91

ASTM (1998)

Type of Test:

Static renewal (every 24 h)

Date:

May 15-19, 2000

Investigator:

S. D. Turley

Laboratory:

UMD/WREC

Chemical:

Designation:

U2723

Source:

Wildlife International, Ltd.

General Water Quality:

See Table A1-1

Test Medium:

FETAX solution

Test Organism:

Scientific Name:

Xenopus laevis

Age at Start of Test:

Stage 8 blastula to stage 11 gastrula

Source:

UMD/WREC culture

Experimental Chambers:

Material:

Glass petri dishes

10 mL

Test Solution Volume:
No. Organisms/Replicate:

25

No. Organisms/Treatment:

Control: 100

Treatment: 50

Lighting:

Fluorescent; 60-85 foot candles

Endpoints:

Mortality; malformation; growth

Test Temperature:

24 ± 2°C

Results:

Note: For Assay 1, statistical calculations are based on the mean measured concentrations of 2.00, 2.83, 4.73, 7.90, 14.70 and 24.60 mg a.i./L, rather than on the nominal concentrations.

Mortality:

The 96-h LC50 = 13.8 mg a,i./L (95% confidence limits = 12.38 - 15.31).

The mortality data are summarized in Table A1-2.

Malformations:

The 96-h EC50 (malformed embryos) = 12.1 mg a.i./L (95% confidence limits = 10.00 - 14.58).

The number of malformed embryos are summarized in Table A1-2. The types of malformations are summarized in Table A1-3.

Growth:

Chemical U2723 had no effect on embryonic growth at any of the test treatments with surviving embryos. As a result, the minimum concentration inhibiting growth (MCIG) could not be calculated.

The growth data are summarized in Table A1-4.

Teratogenic Index:

The teratogenic index (TI) = 1.1.

Table A1-1. Summary of The General Water Chemistry Data for Chemical U2723 - Assay 1 - Dissolved Oxygen (mg/L)

	Test Concentrations (mg/L)						
ŝ	0.00	1.82	3.07	5.19	8.64	14.40	24.00
Day 0							
0 H	8.3	8.4	8.4	8.4	8.4	8.4	8.5
Day 1							
24 H	8.0	7.8	7.9	7.9	8.0	8.0	7.9
0 H	8.4	8.4	8.5	8.4	8.5	8.5	8.4
Day 2							
24 H	7.7	7.7	7.6	7.7	7.8	7.8	7.8
0 H	8.3	8.3	8.4	8.3	8.2	8.3	8.3
Day 3							
24 H	7.4	7.5	7.7	7.7	7.8	7.9	7.9
он	8.3	8.2	8.1	8.2	8.1	8.1	8.1
Day 4							
24 H	7.7	7.7	7.6	7.5	7.7	7.7	7.7
Mean	8.1	8.0	8.1	8.1	8.1	8.1	8.1
Min	7.4	7.5	7.6	7.7	7.8	7.8	7.8
Max	8.4	8.4	8.5	8.4	8.5	8.5	8.5

Table A1-1. (Continued) - pH (Standard Units)

	Test Concentrations (mg/L)										
	0.00	1.82	3.07	5.19	8.64	14.40	24.00				
Day 0			**************************************	1000000			Walls				
0 H	7.74	7.78	7.79	7.78	7.78	7.76	7.62				
Day 1											
24 H	7.26	7.24	7.30	7.34	7.35	7.34	7.33				
OН	7.24	7.45	7.38	7.43	7.43	7.44	7.32				
Day 2											
24 H	7.16	7.17	7.20	7.23	7.23	7.23	7.25				
0 H	7.22	7.25	7.28	7.32	7.34	7.35	7.27				
Day 3											
24 H	7.11	7.13	7.16	7.17	7.25	7.18	6.95				
0 H	7.13	7.20	7.23	7.25	7.26	7.26	7.18				
Day 4											
24 H	7.11	7.15	7.18	7.19	7.18	7.20	7.19				
Min	7.11	7.13	7.16	7.17	7.18	7.18	6.95				
Max	7.74	7.78	7.79	7.78	7.78	7.76	7.62				

Table A1-1. (Continued) - Temperature (°C)

70	Test Concentrations (mg/L)									
	0.00	1.82	3.07	5.19	8.64	14.40	24.00			
Day 0										
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
Day 1										
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
Day 2										
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
Day 3										
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
Day 4										
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
Mean	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
Min	24.0	24.0	24.0	24.0	24.0	24.0	24.0			
Max	24.0	24.0	24.0	24.0	24.0	24.0	24.0			

Table A1-2. FETAX Toxicity Test Data - Percent Embryo Mortality and Malformations after 96 Hours of Exposure to Chemical U2723 - Assay 1

	1		
W	A	0	()
4 .	1	A	W

Concentration (mg/L)	Rep	Number Embryos Alive	Percent Mortality	Number Embryos Malformed	Percent Malformed
Control	1	24	4.0	1	4.2
	2	25	0.0	1	4.0
	3	25	0.0	1	4.0
	4	25	0.0	1	4.0
1.82	1	24	4.0	2	8.3
	2	25	0.0	2	8.0
3.07	1	24	4.0	2	8.3
	. 2	24	4.0	5	20.8
5.19	1	23	8.0	7	30.4
	2	22	12.0	3	13.6
8.64	1	21	16.0	4	19.0
	2	23	8.0	7	30.4
14.40	1	16	36.0	9	56.3
	2	15	40.0	11	73.3
24.00	1	0	100		
	2	0	100		

Table A1-3. FETAX Toxicity Test Data - Type and Number of Malformed Embryos after 96 Hours Exposure to Chemical U2723 - Assay 1

Melformation							Test	Concen	bretion (mg/L)						
	0 Rep			1.82 Rep		3.07 Rep		5.19 Rep		8.64 Rep		14.40 Rep		24.00 Rep		
	1	2	3	4	1	2	1	2	1	2	1	2	1	2	1	2
Severe												1				
Stunted											ŝ					
Gut	1	1	1	1	1	1	2	5	7	3	4	5	7	11		
Edema:																
Multiple								2	2		2	2	4	3		
Cardiac		1					1	3	1		2	2	4	1		
Abdominal							1	3	1		2	1	3	2		
Facial								2	2		2	2	4	2		
Cephalic																
Optic								2	2		1	1	2	6		
Tall																
Notochord												1	1			
Fin																
Face	1	1		1	2	2	1	1	4	3	2	3	4	6		
Eye												1	1			
Bruin																
Нетоппаде																
Cardiac						1			2				3	1		
Blisters																
Other*																
# Malformed	1	1	1	1	2	2	2	5	7	3	4	7	9	11		

^{*} Anal edema.

Table A1-4. FETAX Toxicity Test Data - Embryonic Growth (Length) after 96 Hours Exposure to Chemical U2723 - Assay 1

Concentration (mg/L)	Rep	Number Embryos Alive	Average Length (mm)	Mean Average Length (mm)
Control	1	24	9.05	8.59
	. 2	25	8.40	
	3	25	8.29	
	4	25	8.62	
1.82	1	24 .	8.44	8.29
	2	25	8.14	
3.07	1	24	8.88	8.80
	2	24	8.72	
5.19	1	23	8.42	8.51
	2	22	8.60	
8.64	1	21	8.83	8.71
	2	23	8.58	
14.40	1	16	8.41	8.08
	2	15	7.74	
24.00	° 1	0		
	2	0		

APPENDIX 2

FROG EMBRYO TERATOGENESIS ASSAY-XENOPUS (FETAX) CONDUCTED WITH CHEMICAL U2723 (ASSAY 2)

Test Method:

ASTM Designation E 1439-91

ASTM (1998)

Type of Test:

Date:

Static renewal (every 24 h)

May 22-26, 2000

Investigator:

S. D. Turley

Laboratory:

UMD/WREC

Chemical:

Designation:

U2723

Source: General Water Quality: Wildlife International, Ltd.

See Table A2-1

Test Medium:

FETAX solution

Test Organism:

Scientific Name:

Xenopus laevis

Age at Start of Test:

Stage 8 blastula to stage 11 gastrula

Source:

UMD/WREC culture

Experimental Chambers:

Material:

Glass petri dishes

Test Solution Volume:

10 mL

No. Organisms/Replicate:

25

No. Organisms/Treatment:

Control: 100 Treatment: 50

Lighting:

Fluorescent; 60-85 foot candles

Endpoints:

Mortality; malformation; growth

Test Temperature:

24 ± 2°C

Results:

Note: For Assay 2, statistical calculations are based on the mean measured concentrations of 1.91, 3.04, 4.82, 7.97, 13.30 and 23.10 mg a.i./L, rather than on the nominal concentrations.

Mortality:

The 96-h LC50 = 17.6 mg a.i./L (95% confidence limits = 15.53 -19.96).

The mortality data are summarized in Table A2-2.

Malformations:

The 96-h EC50 (malformed embryos) = 17.6 mg a.i./L (95% confidence limits = 13.49 - 22.88)

The number of malformed embryos are summarized in Table A2-2. The types of malformations are summarized in Table A2-3.

Growth:

The minimum concentration inhibiting growth (MCIG) =7.97 mg a.i./L.

The growth data are summarized in Table A2-4.

Teratogenic Index:

The teratogenic index (TI) = 1.0.

Table A2-1. Summary of The General Water Chemistry Data for Chemical U2723 - Assay 2 - Dissolved Oxygen (mg/L)

			Test Co	ncentration	s (mg/L)		
	0.00	1.82	3.07	5.19	8.64	14.40	24.00
Day 0						100000	
0 H	8.2	8.2	8.1	8.2	8.1	8.2	8.2
Day 1							
24 H	8.3	8.3	8.0	8.1	8.1	8.1	8.2
0 H	8.2	8.2	8.2	8.2	8.1	8.1	8.2
Day 2							
24 H	7.6	7.8	7.5	7.2	7.2	7.2	7.1
0 H	8.1	8.1	8.0	8.1	8.0	8.0	8.1
Day 3				(MAX)	3.0	120	
24 H	7.5	7.6	7.5	7.6	7.7	7.3	7.2
0 H	8.1	8.2	8.1	8.1	8.0	8.1	8.0
Day 4							
24 H	7.6	7.6	7.5	7.5	7.6	7.5	7.5
Mean	8.0	8.0	7.9	7.9	7.9	7.8	7.8
Min	7.5	7.6	7.5	7.2	7.2	7.2	7.1
Max	8.3	8.3	8.2	8.2	8.1	8.2	8.2

Table A2-1. (Continued) - pH (Standard Units)

			Test Co	ncentration	s (mg/L)		
	0.00	1.82	3.07	5.19	8.64	14.40	24.00
Day 0							
0 H	7.14	7.24	7.31	7.33	7.37	7.40	7.37
Day 1							
24 H	7.12	7.17	7.21	7.23	7.24	7.26	7.27
0 H	7.11	7.21	7.28	7.35	7.35	7.39	7.34
Day 2							
24 H	7.18	7.22	7.23	7.24	7.24	7.23	7.24
0 H	7.15	7.23	7.30	7.37	7.38	7.41	7.32
Day 3							
24 H	7.20	7.20	7.22	7.25	7.24	7.23	7.25
0 H	7.17	7.21	7.26	7.34	7.36	7.40	7.35
Day 4							
24 H	7.11	7.21	7.20	7.29	7.33	7.39	7.34
Min	7.11	7.17	7.20	7.23	7.24	7.23	7.24
Max	7.20	7.24	7.31	7.37	7.38	7.41	7.37

Table A2-1. (Continued) - Temperature (°C)

			Test Co	ncentration	s (mg/L)		
	0.00	1.82	3.07	5.19	8.64	14.40	24.00
Day 0							
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Day 1							
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Day 2							
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Day 3						¥.	
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Day 4							
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Mean	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Min	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Max	24.0	24.0	24.0	24.0	24.0	24.0	24.0

Table A2-2. FETAX Toxicity Test Data - Percent Embryo Mortality and Malformations after 96 Hours of Exposure to Chemical U2723 - Assay 2

					*
Concentration (mg/L)	Rep	Number Embryos Alive	Percent Mortality	Number Embryos Malformed	Percent Malformed
Control	1	25	0.0	1	4.0
	2	25	0.0	2	8.0
	3	25	0.0	0	0.0
¥	4	24	4.0	1	4.2
1.82	1	25	0.0	2	8.0
	2	25	0.0	2	8.0
3.07	1	23	8.0	2	8.7
	2	22	12.0	0	0.0
5.19	1	24	4.0	1	4.2
	2	22	12.0	4	18.2
8.64	1	22	12.0	4	18.2
	2	23	8.0	5	21.7
14.40	1	22	12.0	4	18.2
	2	13	48.0	9	69.2
24.00	1	9	64.0	6	66.7
	2	6	76.0	4	66.7

Table A2-3. FETAX Toxicity Test Data - Type and Number of Malformed Embryos after 96 Hours Exposure to Chemical U2723 - Assay 2

Malformation							Test (Concen	ration (mg/L)						
		() ep			62 ep	3.0 Re		5.º		8.0 Re		14. R	40	24.00 Rep	
0	1	2	3	4	1	2	1	2	1	2	1	2	1	2	1	2
Severe																
Stunted																
Gut		1			1	1	1			4	1	3	2	6	6	4
Edema:																
Multiple					2					2		1			3	
Cardiac					2					2	2	1			4	
Abdominal					2					3		1	1		3	
Facial					2				1	3		1			3	
Cephalic																
Optic					2					1					3	
Tell																
Notochord				1		1	2						1	1		
Fin																
Face	1	2				1				2	2	3	1	9	1	;
Eyo																
Brain														¥		
Hemorrhage																
Cardiac		2														
Blisters																
Other*											1			2		
# Malformed	1	2	0	1	2	2	2	0	1	4	4	5	4	9	6	

⁶ Anal edema.

Table A2-4. FETAX Toxicity Test Data - Embryonic Growth (Length) after 96 Hours Exposure to Chemical U2723 - Assay 2

Concentration (mg/L)	Rep	Number Embryos Alive	Average Length (mm)	Mean Average Length (mm)
Control	1	25	8.94	8.88
	2	25	8.81	
	3	25	8.90	
	4	24	8.88	
1.82	1	25	8.39	8.45
	2	25	8.50	
3.07	1	23	8.42	8.57
	2	22	8.71	
5.19	1	24	8.69	8.72
	2	22	8.74	
8.64	1	22	7.94	7.93*
	2	23	7.92	
14.40	1	22	7.89	7.51*
	2	13	7.13	
24.00	1	9	7.50	7.39*
	2	6	7.27	

^{*} Significantly different at alpha = 0.05 (Bonferroni T-Test)

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APPENDIX 3

FROG EMBRYO TERATOGENESIS ASSAY-XENOPUS (FETAX) CONDUCTED WITH CHEMICAL U2723 (ASSAY 3)

Test Method:

ASTM Designation E 1439-91

ASTM (1998)

Type of Test:

Static renewal (every 24 h)

Date:

May 22-26, 2000

Investigator.

S. D. Turley

Laboratory:

UMD/WREC

Chemical:

Designation:

U2723

Source:

Wildlife International, Inc.

General Water Quality:

See Table A3-1

Test Medium:

FETAX solution

Test Organism:

Scientific Name:

Xenopus laevis

Age at Start of Test:

Stage 8 blastula to stage 11 gastrula

Source:

UMD/WREC culture

Experimental Chambers:

Material:

Glass petri dishes

Test Solution Volume:

10 mL _

No. Organisms/Replicate:

25

No. Organisms/Treatment:

Control: 100

Treatment: 50

Lighting:

Fluorescent; 60-85 foot candles

Endpoints:

Mortality; malformation; growth

Test Temperature:

24 ± 2°C

Results:

Note: For Assay 3, statistical calculations are based on the mean measured concentrations of 1.93, 3.27, 5.25, 8.26, 14.00 and 23.90 mg a.i./L, rather than on nominal concentrations.

Mortality:

The 96-h LC50 = 15.3 mg a.i./L (95% confidence limits = 13.13 - 17.75).

The mortality data are summarized in Table A3-2.

Malformations:

The 96-h EC50 (malformed embryos) = 16.8 mg a.i./L (95% confidence limits = 12.35 - 22.82).

The number of malformed embryos are summarized in Table A3-2. The types of malformations are summarized in Table A3-3.

Growth:

The minimum concentration inhibiting growth (MCIG) =8.26 mg a.i./L.

The growth data are summarized in Table A3-4.

Teratogenic Index:

The teratogenic index (TI) = 0.9.

Table A3-1. Summary of The General Water Chemistry Data for Chemical U2723 - Assay 3 - Dissolved Oxygen (mg/L)

	Company of Control		Test Co	ncentration	s (mg/L)		
	0.00	1.82	3.07	5.19	8.64	14.40	24.00
Day 0							
0 H	8.2	8.2	8.1	8.2	8.1	8.2	8.2
Day 1							
24 H	8.3	8.3	8.0	8.2	8.2	8.2	8.3
0 H	8.2	8.2	8.2	8.2	8.1	8.1	8.2
Day 2							
24 H	7.4	7.2	7.1	7.0	7.0	7.0	7.0
0 H	8.1	8.1	8.0	8.1	8.0	8.0	8.1
Day 3							
24 H	7.3	7.1	7.1	7.0	7.0	7.1	7.0
0 H	8.1	8.2	8.1	8.1	8.0	8.1	8.0
Day 4							
24 H	7.4	7.5	7.5	7.5	7.6	7.4	7.4
Mean	7.9	7.9	7.8	7.8	7.8	7.8	7.8
Min	7.3	7.1	7.1	· · 7. 0	7.0	7.0	7.0
Max	8.3	8.3	8.2	8.2	8.2	8.2	8.3

Table A3-1. (Continued) - pH (Standard Units)

			Test Co	ncentration	s (mg/L)		
	0.00	1.82	3.07	5.19	8.64	14.40	24.00
Day 0							
0 H	7.14	7.24	7.31	7.33	7.37	7.40	7.37
Day 1							
24 H	7.18	7.19	7.15	7.14	. 7.15	7.14	7.16
0 H	7.11	7.21	7.28	7.35	7.35	7.39	7.34
Day 2							
24 H	7.21	7.20	7.18	7.17	7.17	7.15	7.23
0 H	7.15	7.23	7.30	7.37	7.38	7.41	7.32
Day 3		•					
24 H	7.23	7.21	7.25	7.26	7.24	7.28	7.25
0 H	7.17	7.21	7.26	7.34	7.36	7.40	7.35
Day 4							
24 H	7.18	7.21	7.24	7.31	7.35	7.40	7.29
Min	7.11	7.19	7.15	7.14	7.15	7.14	7.16
Max	7.23	7.24	7.31	7.37	7.38	7.41	7.37

Table A3-1. (Continued) - Temperature (°C)

			Test Co	ncentration	s (mg/L)		
	0.00	1.82	3.07	5.19	8.64	14.40	24.00
Day 0							
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Day 1							
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Day 2							
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Day 3							
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
0 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Day 4							
24 H	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Mean	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Min	24.0	24.0	24.0	24.0	24.0	24.0	24.0
Max	24.0	24.0	24.0	24.0	24.0	24.0	24.0

Table A3-2. FETAX Toxicity Test Data - Percent Embryo Mortality and Malformations after 96 Hours of Exposure to Chemical U2723 - Assay 3

Concentration (mg/L)	Rep	Number Embryos Alive	Percent Mortality	Number Embryos Malformed	Percent Malformed
Control	1	25	0.0	1	4.0
	2	25	0.0	1	4.0
	3	25	0.0	0	0.0
	4	25	0.0	0	0.0
1.82	1	25	0.0	*1	4.0
	2	25	0.0	2	8.0
3.07	1	25	0.0	1	4.0
	2	25	0.0	°Ĵ	4.0
5.19	1	25	0.0	1	4.0
	2	25	0.0	2	8.0
8.64	1	23	8.0	1	4.3
	2	20	20.0	5	25.0
14.40	1	14	44.0	4	28.6
	2	14	44.0	7	50.0
24.00	1	6	76.0	5	83.3
receives a see Time Time	2	5	80.0	3	60.0

Table A3-3. FETAX Toxicity Test Data - Type and Number of Malformed Embryos after 96 Hours Exposure to Chemical U2723 - Assay 3

Matformation		***				ASSES A	Test (Concent	ration (mg/L)						
:	Ess		0 lep		1.0 Re		3.0 Re		5.1 Re		8.0 Re		14. Re			.00 ep
	1	2	3	4	1	2	1	2	1	2	1	2	1	2	1	2
Sovere				***												
Stunted																
Gut		1				2		1			1	3			2	3
Edoma:																
Multiple											1				280	
Cardino											1		•	3		
Abdominal											1					
Facial											1					
Cephalic					54											
Optic							6,	•			1					
Tell																
Notochord						1				1		3	3		1	
Fin a																
Fece	1				1	1	1		1	1		3	3	4	3	;
Eye																
Brain																
Hemorrhage																
Cardiac																2
Blisters																
Other*					4.10	1	32.3					3	1	3	1	
# Malformed	1	1	0	0	1	2	1	1	1	2	1	5	4	7	5	;

^{*} Anal edema.

Table A3-4. FETAX Toxicity Test Data - Embryonic Growth (Length) after 96 Hours Exposure to Chemical U2723 - Assay 3

Concentration (mg/L)	Rep	Number Embryos Alive	Average Length (mm)	Mean Average Length (mm)
Control	1	25	9.54	9.47
	2	25	9.64	
	3	25	9.31	
	4	25	9.40	
1.82	1	25	8.83	9.10
99	2	25	9.36	
3.07	1	25	9.11	9.28
	2	25	9.44	
5.19	1	25	9.25	9.28
	2	25	9.30	
8.64	1	23	8.70	8.51°
	2	20	8.31	
14.40	1	14	8.08	8.11*
	2	14	8.14	
24.00	1	6	8.06	7.80*
	2	5	7.53	

^{*} Significantly different at alpha = 0.05 (Bonferroni T-Test)

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APPENDIX 4

ASSAY 1 RAW DATA SHEETS FOR FROG EMBRYO TERATOGENESIS ASSAY-XENOPUS (FETAX) AND 6-AMINONICOTINAMIDE REFERENCE TOXICANT FOR CHEMICAL U2723

Note: The nominal concentrations of 1.9, 3.2, 5.4, 9.0, 15.0 and 25.0 mg/L in the raw data correspond to the nominal concentrations of 1.82, 3.07, 5.19, 8.64, 14.40 and 24.00 mg a.i./L in the report.

FORM 8D (Page 1 of 2): FETAX TOTAL SUMMARY DATA SHEET Munchook/Binder Mu.____

Concentration Embryos Marally	N Tel	H	Table of Smbone				í	
	_	Mortality	Alive at 96 hrs.	Malformed	Malformed	Mattermed Mattermed	Average Length	Mean Average Length (mm)
Control A 25 1	4	-	24	-	4.2	4.0	9,05	8,59
8 25			প্র	. 1	4.0		8.40	
52	0		SS		4.0		8.29	
Colle D 25 0	0		35	-	40		8.62	
8	4	.2	74	2	8.3	8.2	8.44	8.29
8			32	2	8.0		8.14	
V	. 4	4	24	2	8.3	14.6	88.8	08.80
8	4		4	2	20.8		8.72	
¥	Œ	0	ಬ	7	30.4	22.0	8.42	8.51
8 22	12		22	3	13.6		8.6	
83 V	9	12	. 12	4	19.0	2450	883	P 00
8	ထ		23	7	30.4		8 58	
4	%	38	91	6	56.3	648	841	808
8 35			15	=	73,3		7.74	
٧	100	001	0					
B 25	100		0					
A 25	8	%) (C)	. 23	5	65.2	55.4	6),8	118
8 25	12		22	0	455		8.15	
A 25	8	.00	0					
250m, R 25 25	8		0					

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FI IKAA	KI .	FFIAX	WE BE LULLE	IIAIA

Page_

Test Number Definitive Tat 1

Test Material U2723

		-		Mort	ality				
Petri Dish/ Aquarium ID	Concentration	Initial No. of Embryos	24 Hr.	48 Hr.	72 Hr.	96 Hr.	Total No. Dead Embryos	% Dead Embryos	Total No. Embryos Alive/Fixed
01	FETTH Control A	25	0	0	0	1	-	4	24
02	FSTAX Control B	25	0	0	0	0	0	0	25
03	FERX Colol C	22	0	0	0	0	0	. 0	25
04	PETAX Control D	25	0	0	0	0	0	0	25
05	1.9 mg/L A	ट्ट	0	0	1.	0	1	4	24
06	1.9 mg/L B	52	0	0	0	0	0	0	25
07	3.2 mg/ A	52	0	0	0	. 1	1	4	24
08	3.2 mg/L B	25	0	0	0	1	1	4	24
09	5.4 mg/L A	52	0	2	0	0	2	8	23
10	5.4 mall B	25	0	. 0	2	_1_	3	12	22
11	9.0 mg/L A	25	2	2	0	0	4	16	21
12	90mil B	25	0	2	0	0	2	8	23
13	15.0mll A	52	0	2	7	0	9	36	16
14	ISDAL B		2		5	2	10	40	15
15	250mll A	52	0	2	19	4	25	100	0
16	Somit B	25	0	6	17	2	25	100	0
			$\downarrow =$					1	
			 			├─	-		<u> </u>
	 	-	-		+-	-	+	+	-
	1								
						1			
L		By SX	BYST	BYSX	107 ₅ X	8757	+		By €I
		Date S	Date 5 %	Date 5/17	S B	SIA			Date 5/19

2.7. 12.	0
Raviewed By	 Oate





FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Data Collected By Ster Terley Date 5/15-6/19/00 Tost Mumber Definiting Took # Test Malerial U2723 Concentration FETTA Control Petri Dish (Mai) No. Ol

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FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Page_ Date 5/15-5/19/00 Test Material 112723 CONCENTION FETAX Control B Petri Dish (Mal) Ha. 02.

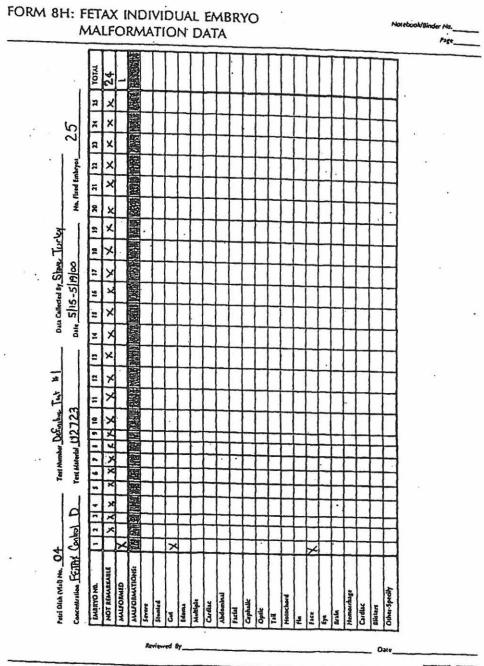


FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA · 00/6/15-5/15 MILE Concentration PETRIA Control Peul Dish (Mul) No. 03

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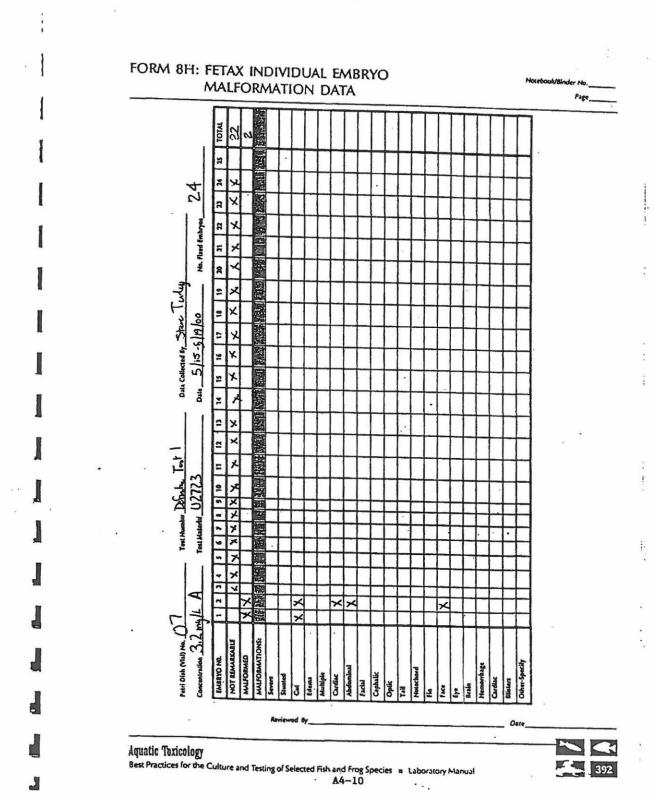
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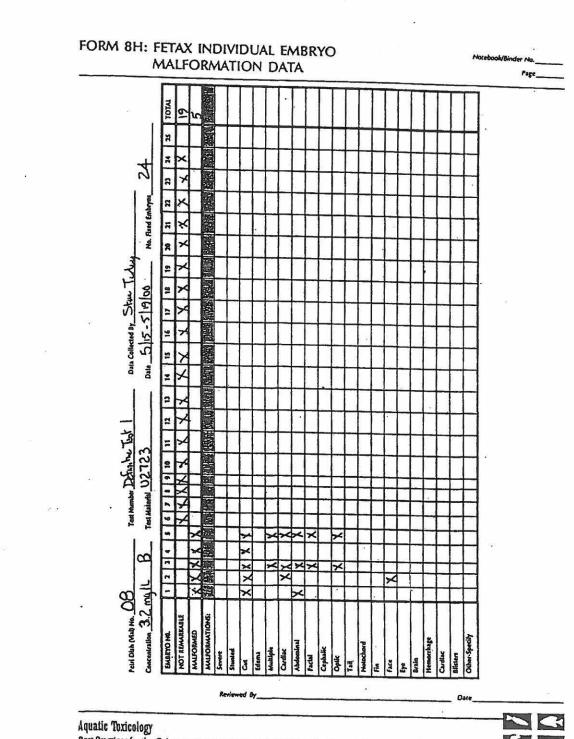
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FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Page_ Data Collected By Skue Date 5/16-5/19/00 Test Malerial U2723 1.9 mg/L

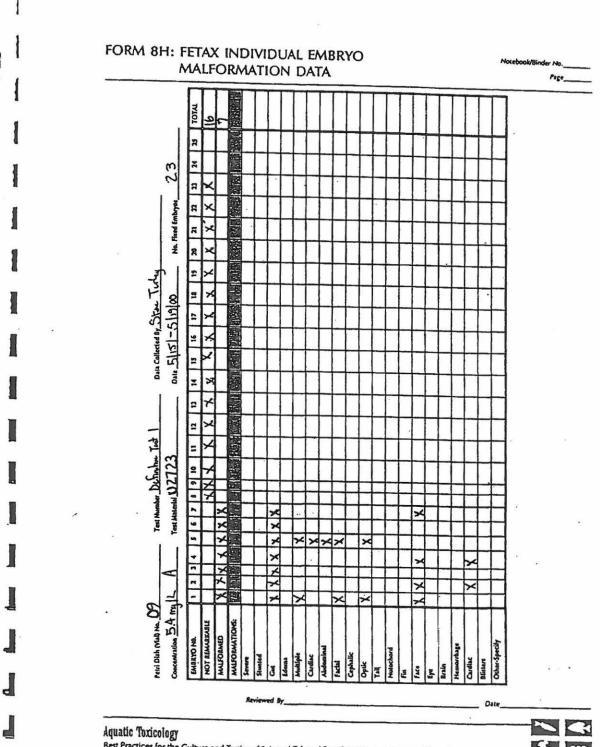
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FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Page_ Date 5/15-6/19/00 Tel Maledia U2723 Concentration 1.9 ms/L





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FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Date 5/15-5/19/00 8 5.4 m Aquatic Toxicology Best Practices for the Culture and Testing of Selected Fish and Frog Species

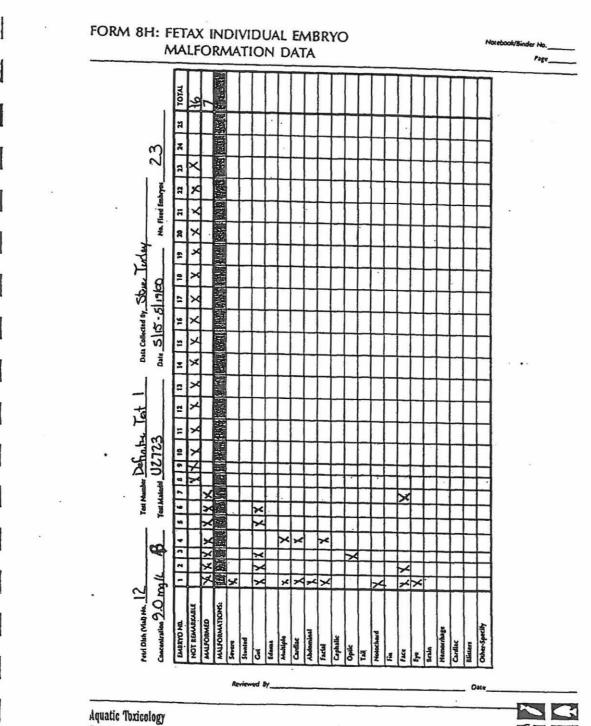
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FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Page_ Date 5/15-5/19/00 72 9.0 mll

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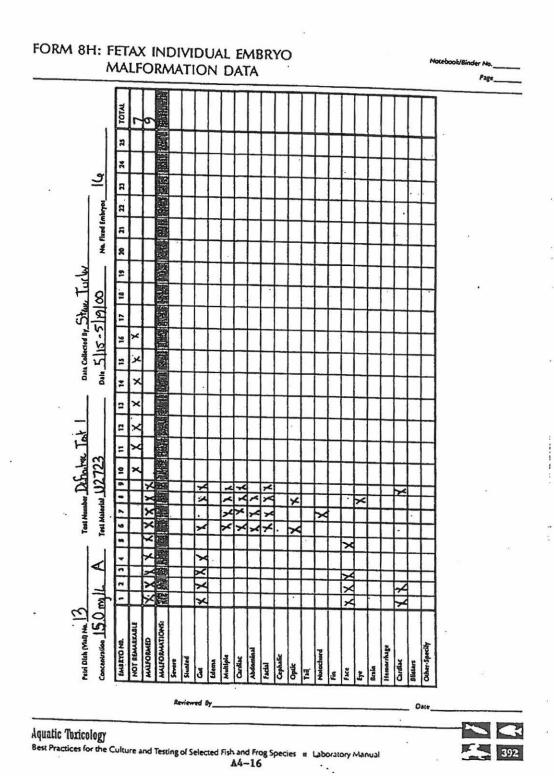
Best Practices for the Culture and Testing of Selected Fish and Frog Species

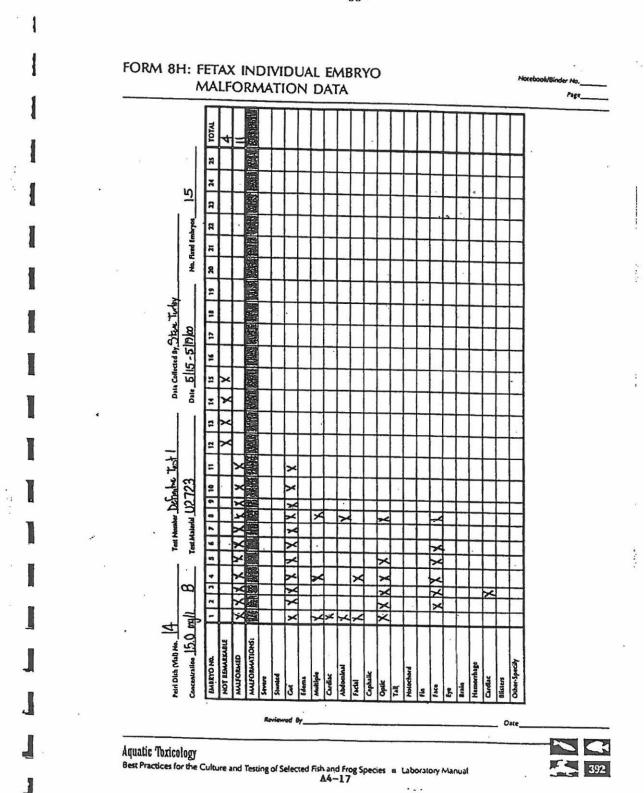
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96 Hour 7	Fest Data	
Effluent/Groundwater/Toxicant) Chemical	U2723	Date: May 15-19,2000
Definitive Test #1		Analyst: Steve Turley
Species: Xenopus Jaevis	Age at Start	of Test: Rbsklo (Stege 8-11)

Control	· Day 0	Da	y 1	Da	y 2	Da	y 3	Day 4
FETAX Control	0·H	Old ·	New	Old	New	Óld	New	Old .
Temperature (°C)	24.0	24,0	240	24.0	24.0	240	24.0	24,0
D.O. (mg/L)	8.3	8.0	8.4	7.7	8.3	7.4	8.3	7.7
рН .	7.74	7.26	7.24	7.16	7.22	7.11	7.13	7.11
Conductivity				1				
Alkalinity .								
Hardness						1		
	1					1	1	
	1	1	1					

1.9 mg/L	Day 0	Day	y 1	Da	y 2	Day	y 3	Day 4
U2723	он .	Old	New	Old	New	Old	New	Old
Temperature (°C)	24.0	24,0	2A.0	24.0	240	240	24.0	24.0
D.O. (mg/L)	8,4	7.8	8.4	7,7	8.3	7,5	8.2	7.7
ρH ·	7.78	7.24	7.45	7.17	7.25	7.13	7.20	7.15
Conductivity			Ι.		[}		
Alkalinity								
Hardness		1		1.				1
		1					1	
	1		1		7			

3.2 mg/L	Day 0	Da	y 1	Da	y 2	Da	y 3	Day 4
UZTZ3	O.H.	Old	New	Old	·N≅w -	Old	New	Old
Temperature (°C)	24.0	24.0	24.0	24.0	24.0	24.0	24,0	24.0
D.O. (mg/L)	8.4	7.9	8.5	7.6	8.4	7,7	8.1	7.6
ρН	7.79	7.30	7.38	7.20	7.28	7.16	7.23	7.18
Conductivity						*		
Alkalinity					T			
Hardness								
	1	1		1	1			

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Definite Test 1 96 Hour Test Data Page 2 of 3

5.4 mg/L	Day 0	Da	y 1	- Da	y 2	Da	y 3	Day 4
U2723	0 H	Old	New	Old	New	Old	New	Old
Temperature (°C)	24.0	24.0	24.0	24D	24.0	24,0	24.0	24.0
D.O. (mg/L)	8.4	7.9	8.4	7.7	8.3	7.7	8.2	7.5
ρΗ	7.78	7.34	7,43	7.23	7.32	7.17	7.25	7.19
Conductivity					1			
Alkalinity			,					
Hardness								
					Ì			

9.0 mg/L	Day 0	Day	y 1	Da	y 2	Da	у 3	Day 4
U2723	ОН	Old	New	Old	New	Old	New	Old
Temperature (°C)	24.0	24.0	24.0	24.0	24.0	24.0	24,0	24.0
D.O. (mg/L)	8.4	80	8.5	7.8	8.2	7.8	8.1	7.7
рН	7.78	7.35	7.43	7.23	7.34	7,25	7.26	7.18
Conductivity								
Alkalinity						1		
Hardness								
	1							
	1							

15.0 mg/L	Day 0	Day	y 1,	Da	y 2	Da	y 3	Day 4
U2723	ОН	Old	New	Old	New	Old	New	Old
Temperature (°C)	24.0	24.0	24.0	74.0	74.0	24.0	24.0	24.0
D.O. (mg/L)	8.4	80	8.5	7.8	8.3	7.9	8.1	7.7
рН	7.76	7.34	7.44	7.23	7.35	7.18	7.26	7.20
Conductivity								
Alkalinity								
Hardness								
		1	1					
	+	+	1		1			

96 Hour Test Data Page 3 of 3

25.0 mg/L	Day 0	Da	y 1	Da	y 2	Da	y 3	Day 4
	ОН	Old	New	Old	New	Old	New	Old
Temperature (°C)	24.0	24.0	74.0	24.0	24.0	24.0	24.0	24.0
D.O. (mg/L)	8.5	7.9	8.4	7.8	83	7.9	8.1	7.7
pН	7.62	7.33	7.32	7.25	7.27	6.95	7.18	7.19
Conductivity								
Alkalinity			S					3
Hardness								
					-			

	Day 0	Day 1		Day 2		Day 3		Day 4
	σн	Old	New	Old	New	Old	New	Old
Temperature (°C)								
D.O. (mg/L)								
рН								
Conductivity								
Alkalinity								
Hardness							T.	
	1.			1		1		
	1				1			

	Day 0	Day 1		Day 2		Day 3		Day 4
	ОН	Old	New	Old	New	Old	New	Old
Temperature (°C)		1						
D.O. (mg/L)								
рН								
Conductivity							1	
Alkalinity		1						1
Hardness								
	1							
			1	na fi				1

96 Ho	ur Test Data
Effluent/Groundwater/Toxicant: 6-Amir	onvolvement (Text 1) Date: 5/15/-5/19/20
	Analyst; ST
Species: Xeropus Caeus	Age at Start of Test: Soc. 8 - Social

Control	Day 0 Day 1		y 1	Day 2		Day 3		Day 4
5.5 mg/L (6-AN)	ОН	Old	New	Old .	New	Old	New	Old
Temperature (°C)	24.0	24.0	24.0	240	24.0	240	240	24.0
D.O. (mg/L)	8.4	8.0	8,3	7.9	8.3	7.8	8.3	7.6
pН	7.56	7.31	7.27	7.19	7.30	7.16	7.76	7.07
Conductivity								
Alkalinity								
Hardness							'	
		1						
		1			,[.			

2500 mg/L (6-AN)	Day 0	Day 1		Day 2		Day 3		Day 4	
•	ОН	Old	New	Old	New	Old	New	Old	
Temperature (°C)	24.0	24.0	24,0	24.0	24.0	24.0	240	240	
D.O. (mg/L)	8.3	7.9	8.3	7,7	8.3	7.6	8.3	7.4	
pH ·	7.44	7.26	7.20	7.4	7.00	7.10	7.17	7.04	
Conductivity		1			İ				
Alkalinity									
Hardness		1		1					
10000								1	
	,			1.	1			1	

	Day 0	Day 1		Day 2		Day 3		Day 4
	OH	Old	New	Old	New "	Old	New	Old
Temperature (°C)								
D.O. (mg/L)								
pH							T	
Conductivity								
Alkalinity			Τ.					
Hardness								
		T						

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Chemical U2723- Definitive Test 1 Embryo Length Measurements (mm) Page 1 of x 3

	Control A	Control B		Control D	1.9 mg/L A	1.9 mg/L B
1	8.86	8.66	8.96	8.78	8.76	8.48
2	9.06	8.03	8.83	8.91	9	6.26
3	9.36	8.25	8.83	8.01	9.11	8.23
4	9.25	8.69	8.07	8.63	8.25	8.2
5	8.93	8.1	8.44	8.08	8.5	8.75
6	9.32	8.35	9.41	8.61	8.82	8.31
7	9.76	8.4	8.05	8.83	7.44	8.3
8	9.29	7.74	7.51	9.1	8.3	7.83
9	9.37	8.38	7.88	8.94	8.38	7.96
10	9.54	8.68	7.96	9	8.23	8.53
11	9.39	7.29	8.1			8.16
12	9.28	8.28	7.96	7.58	8.58	7.27
13	9.29	8.58	9.05	8.01	7.59	
14	8.76	8.81	8.17	8.33	9.45	8.75
15	8.22	8.36	8.28	8.75	8.51	8.41
16	8.19	8.6	7.45	8.83	8.87	8.25
17	7.93	8.85	7.94	8.4	8.17	8.16
18	9.56	8.72	8.17	8.14	8.77	8.16
19	8.96	8.36	7.34	8.54	8.24	8.03
20	8.51	8.55	10.06	8.78	7.88	8.47
21	9.17	9.02	9.22			
22	9.19	8.16	7.47	8.53	8.47	8.27
23	8.58	7.87	7.81	9.46	7.93	7.67
24	9.45	8.73	8.02	8.29	7.42	8.55
25	i	8.64	8.36	9.12	2	7.67
Mean	9.0508333	8.404	8,2936	8.6172	8.44375	8.1352
Min	7.93	7.29	7.34	7.58	7.42	6.26
Max	9.76	9.02	10.06	9.46	9.45	
Stand. Dev	. 0.4723477	0.3886194	0.6695949	0.429627	0.5323027	0.5190466

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Chemical U2723- Definitive Test 1 Embryo Length Measurements (mm) Page 2 of x 3

	3.2 mg/L A	3.2 mg/L B	5.4 mg/L A	5.4 mg/L B	9.0 mg/L A	9.0 mg/L B
1	9.24	9.22	5.61	9.7	8.73	9.16
2	9.15	8,66	8.41	8.92	8.89	8.52
3	8.84	9.4	6.85	7.33	7.24	9.18
4	7.53	8.78	6.63	9.14	8.4	6.34
5	8.95	8.74	8.59	8.98	8.37	8.6
6	9.21	8.76	7.42	8.25	8.44	8.84
7	8.01	9.09	8.98	8.45	9.51	8.89
8	9.25	8.34	8.73	8.37	9.23	7.41
9	9.54	9.37	8.92	6.25	8.66	8.83
10	9.39	8.44	8.6	8.84	8.87	8.73
11	8.07	8.35	8.39	8.75	6.98	9.26
12	10.08	8.03	7.61	9.26	8.32	9.45
13.	9.2	9.04			8.83	
14	8.84	9.18	9.35	8.79	9.15	9.22
15	8.94	8.76	8.16	8.5	9.5	8.99
16	8.09	8.07	9.1	8.93	9.45	9.01
17	9.38	7.17	9.41	9.11	9.81	9.82
18	8.78	8.97	8.72	9.76	9.09	9.4
19	8.08	9.18	8.91	8.72	8.94	6.95
20	9.66	9.58	9,32	8.33	9.6	
21	8.84	8.27	9.57	8.58	9.52	8.24
22	9.28	8.61	8.79	8.27		9.24
23	7.69	8.23	9	K		6.33
24	9.16	9.04				
25						
Mean	8.8833333	8.72	8.4165217	8.6	8.8347619	8.5826087
Min	7.53	7.17	5.61	6.25	6.98	6.33
Max	10.08	9.58	9,57	9.76	9.81	9.82
Stand. Dev.	0.6501483	0.5448654	0.9830824	0.7544029	0.7235373	0.9891145

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Chemical U2723- Definitive Test 1 Embryo Length Measurements (mm) Page 3 of 3

					6-Aminonicot	inamide
	15 mg/L A	15 mg/L B	25 mg/L A	25 mg/L B	5.5 mg/L A	5.5 mg/L B
1	9	6.55			7.95	6.99
2	9.57	7.85			8.21	7.77
3	7.8	7.65			8.46	7.99
4	7.73	7.76			8.3	8.92
5	7.79	8.9			9.6	8.4
6	9.5	7.28			9.02	7.8
7		7.43			7.46	9.12
8					7.59	9.23
9		8.2			7.7	8.11
10		8.48			8.1	7.55
11		6.51			8.16	8.57
12	10.24				8.01	7.96
13					7.99	8.41
14					9.11	8.69
15					8.81	7.95
16					8.41	8.12
17					8.16	9.04
18			•		7.44	8.27
19					7.39	7.84
20					7.8	7.93
21					8.12	7.15
22					8.91	7.38
23					7.65	
24						
25	i		E;			
Mean	8.40875	7.7390909)		8.1891304	8.145
din	7.14	6.51			7.39	6.99
Иах	10.24	8.9	9		9.6	9.23
Stand, Dev	0.9429519	0.7726895	5 .		0.5832504	0.6140556

FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Page_ Test Material 6- Amircarchamide

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FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Page_ Dala Collected By Star Turky 00/11/2-5/12 DO 0 5.5 ml 20 Peirl Dich (Mal) No.

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TRIMMED SPEARMAN-KARBER METHOD. VERSION 1.5

Mortality - LC50 1 40554 - ESTSU

96 h DATE: 5/15/00 TOXICANT: u2723 SPECIES: xenopus TEST NUMBER: 1 DURATION:

RAW DATA:	Concentration	Number	Mortalities
	(mg/l)	Exposed	
	.00	100	1
	2.00	50	1
	2.83	. 50	2
*	4.73	50	5
	7.90	50	6
	14.70	50	19
20	24.60	50	50

1.01% SPEARMAN-KARBER TRIM:

13.77 SPEARMAN-KARBER ESTIMATES: LC50: 95% LOWER CONFIDENCE: 12.38

15.31 95% UPPER CONFIDENCE:

NOTE: MORTALITY PROPORTIONS WERE NOT MONOTONICALLY INCREASING. ADJUSTMENTS WERE MADE PRIOR TO SPEARMAN-KARBER ESTIMATION. DATE: 5/15/00 TOXICANT : U2723 - 79 -

DURATION:

20

TRIMMED SPEARMAN-KARBER METHOD. VERSION 1.5

U2723 - Assay 1 - Malformations (EC50)

SPECIES: xenopus RAW DATA: Concentration Number Mortalities (mg/l)Exposed .00 99 2.00 49 2.83 48 7 4.73 45 10 7.90 44 11

TEST NUMBER: 1

SPEARMAN-KARBER TRIM: 36.98%

14.70

SPEARMAN-KARBER ESTIMATES: EC50: 12.08
95% LOWER CONFIDENCE: 10.00
95% UPPER CONFIDENCE: 14.58

NOTE: MORTALITY PROPORTIONS WERE NOT MONOTONICALLY INCREASING.
ADJUSTMENTS WERE MADE PRIOR TO SPEARMAN-KARBER ESTIMATION.

TI = LC50/EC50 = 13.77/12.08 = 1.14

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Title: u2723:Assay 1: embryo length data (mg) File: ASSAY1LE. Transform:

NO TRANSFORMATION

Shapiro - Wilk's Test for Normality

D = 0.6683 W = 0.9649

Critical W = 0.8250 (alpha = 0.01 , N = 14) W = 0.8740 (alpha = 0.05 , N = 14)

Data PASS normality test (alpha = 0.01). Continue analysis.

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Title: u2723:Assay 1: embryo length data (mg)
File: ASSAY1LE. Transform:

NO TRANSFORMATION

Bartlett's Test for Homogeneity of Variance

Calculated B1 statistic = 2.3953

(p-value = 0.7922)

Data PASS B1 homogeneity test at 0.01 level. Continue analysis.

Critical B = 15.0863 (alpha = 0.01, df = 5) = 11.0705 (alpha = 0.05, df = 5)

Using Average Degrees of Freedom (Based on average replicate size of 2.33)

Calculated B2 statistic = 2.2190

(p-value = 0.8181)

Data PASS B2 homogeneity test at 0.01 level. Continue analysis.

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Title: u2723:Assay 1: embryo length data (mg) File: ASSAY1LE. Transform:

NO TRANSFORMATION

ANOVA Table

SOURCE	DF	SS	MS	F
Between	5	0.7451	0.1490	1.7838
Within (Error)	8	0.6683	0.0835	
Total	13	1.4134		

(p-value = 0.2224)

Critical F = 6.6318 (alpha = 0.01, df = 5,8) = 3.6875 (alpha = 0.05, df = 5,8)

Since F < Critical F FAIL TO REJECT Ho: All equal (alpha = 0.05)

Title: u2723:Assay 1: embryo length data (mg)
File: ASSAY1LE. Transform:

NO TRANSFORMATION

` Bo	onferroni t-Test -	TABLE 1 OF 2	Ho: Control	L <treatme< th=""><th>nt</th></treatme<>	nt
GROUP	IDENTIFICATION	TRANSFORMED MEAN	MEAN CALCULATED IN ORIGINAL UNITS	t STAT	SIG 0.05
					-
1	con	8.5900	8.5900		
2	2.00 mg/l	8.2900	8.2900	1.1985	
3	2.83 mg/l	8.8000	8.8000	-0.8390	
4	4.73 mg/l	8.5100	8.5100	0.3196	
5	7.90 mg/l	8.7050	8.7050	-0.4594	
6	14.70 mg/l	8.0750	8.0750	2.0575	

Bonferroni t critical value = 2.8965 (1 Tailed, alpha = 0.05, df = 5,8)

Title: u2723:Assay 1: embryo length data (mg) File: ASSAY1LE. Transform:

NO TRANSFORMATION

1	Bonferroni t-Test -	TABLE 2	OF 2	io: Control	L <treatment< th=""></treatment<>
GROUP	IDENTIFICATION	NUM OF REPS	MIN SIG DIFF (IN ORIG. UNITS)	% OF CONTROL	DIFFERENCE FROM CONTROL
1	con	4			
2	2.00 mg/l	2	0.7250	8.4	0.3000
3	2.83 mg/l	2	0.7250	8.4	-0.2100
4	4.73 mg/l	2	0.7250	8.4	0.0800
5	7.90 mg/l	2	0.7250	8.4	-0.1150
6	14.70 mg/l	2	0.7250	8.4	0.5150

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APPENDIX 5

ASSAY 2 RAW DATA SHEETS FOR FROG EMBRYO TERATOGENESIS ASSAY-XENOPUS (FETAX) AND 6-AMINONICOTINAMIDE REFERENCE TOXICANT FOR CHEMICAL U2723

Note: The nominal concentrations of 1.9, 3.2, 5.4, 9.0, 15.0 and 25.0 mg/L in the raw data correspond to the nominal concentrations of 1.82, 3.07, 5.19, 8.64, 14.40 and 24.00 mg a.i./L in the report.

Total Na. of Embryos Aliva 136 har. Malformed 25 2 25 0 25 0 25 0	Mulfbrmed	Malformed A	Length Length	Mesa Average Length
-00-	4.0	17	A 94	
70-				88.88
ю <i>-</i>	.00		8,81	
_	0		8.30	
-	4.2		8.98	
2	8,0	8.0	€'8	8.45
2	8.0		8.50	
2	8.7	44	8.42	850
0	0		11.8	
-	4.2	11.2	8.69	8.72
	18.2		8.74	
4	18.2	20.0	7.94	793
	21.7		792	
	18.2	43.7	7.89	7.51
	69.2		7.13	
	1,099	1,003	7.50	7.39
	66.7		7.27	
12	50.0	-58	7.41	7.35
1 14 (1.09		7.28	
			·	
	-			
-			-	
			69.2 69.2 66.7 66.7 66.1 66.1 66.1 66.1 66.1	18.2 437 69.2 66.7 66.7 66.7 66.1 66.1 66.1

Aquatic Toxicology

Best Practices for the Culture and Testing of Selected Fish and Frog Species Laboratory Manual

A5-2



NAGO:	ac.	EETAV	MORTAL	ITV I	ATA

Notebookil	Binder No.	
	Page	7

		ſ		Morta	ılity		*		
Petri Dish/ Aquarium ID	Concentration	Initial No. of Embryos	24 Hr.	48 Hr.	72 Hr.	96 Hr.	Total No. Dead Embryos	% Dead Embryos	Total No. Embryos Alive/Fixed
01	FETAGGOND A	25	0	0	0	0	0	0	25
OZ	PETAKCOLD B	25	0	0	0	0	0	0	25
03	FETTANCOLLIC	25	0	0	0	0	0	0	25
04	FETAL CILLID	25	0	0	0	_1		4.0	24
02	1.9mg/L A	চ	0	0	0	0	0	0	25
06	1.9m/L B	25	0	0	0	_0_	0	0	25
70	3.2m/L A	22	0	2	0	0	2	8	23
08	3.2 mg/L B	25	0		0	2	3	12	22
٥٩	5.4 mgl A	25	6	0	0	i	1	4	24
10	54m/L B	25	0		80	2	3	12	22
	9.0m/L A	25	0		0	2	3_	12	22
12	9.0 mg/L B	25	0	1	0	_1_	2	8	23
13	KO TIL A	25			0	7	3	12	22
14	15.0 ml B	25_	0	3	3	6	12	48	13
15	25.0ml A	25	0		13	2	16	64	9
16	250mjl B	25	2	$\vdash \vdash \vdash$. Ile	5	19	76	6
	6-Amusadind								
19	5.5 mgl A	25	0	0	1	0	1.1	4	24
20	55mil B	25	0	0	1	1.	2	8	23
21	2500 myle A	25	9	9	8	8_	25	100	0
22	2500my/L B	25	4	8	88	5	2.5	100	0
	ļ								
		By SUTE	8y ST	Bγ ≤₹	BYST	°7ST	- 11 (A 14 (A (A 14 (A (A)(A)(A)(A)(A)(A)(A)(A)(A)(A)(A)(A)(A)(<u></u>	T8 ^{Va}
		Date 500	Date 5/23	°5724	5 25	Date 5 26]		Oate 5/24

Aquatic Toxicology

Best Practices for the Culture and Testing of Selected Fish and Frog Species at Laboratory Manual A5-3



FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Dale 5/22-5/26/00 Concentration FETAX Control Petri Dich (Mal) No. OI

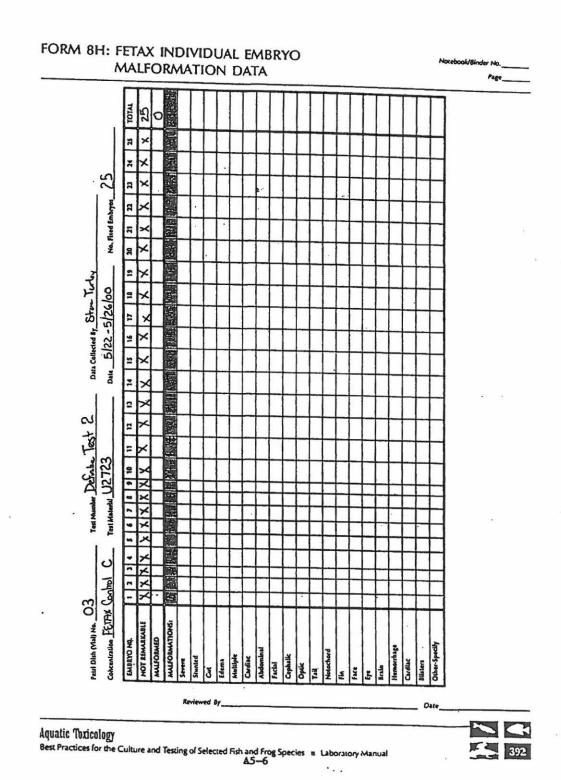
Aquatic Toxicology

Best Practices for the Culture and Testing of Selected Fish and Frog Species

Laboratory Manual

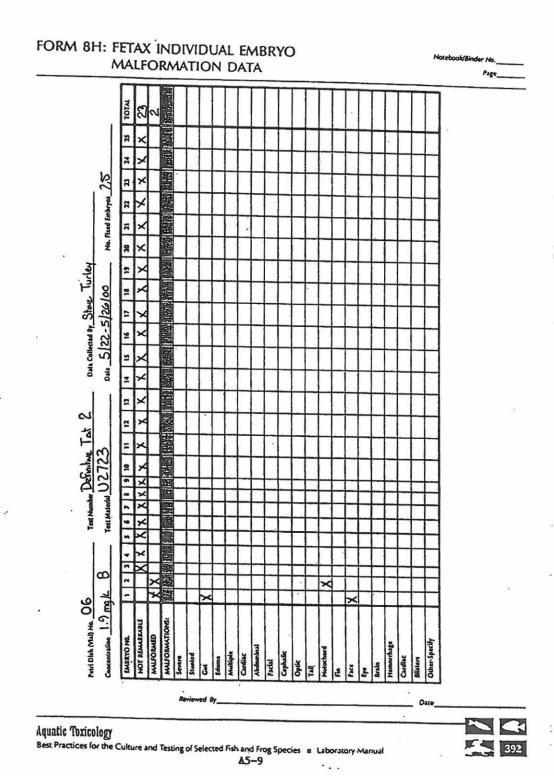


FORM 8H: FETAX INDIVIDUAL EMBRYO Notebook/Binder No. MALFORMATION DATA Page_ R Date 5/22-5/26/00 Concentration FETTY Control Peri Dish (Mai) No. 02 Aquatic Toxicology



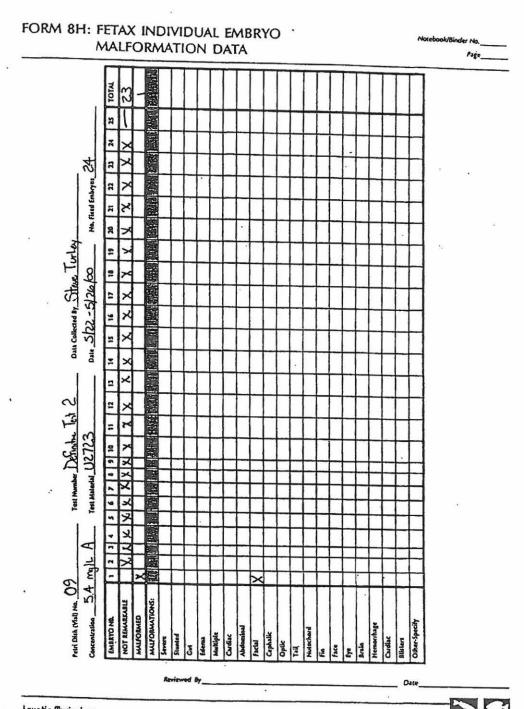
FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA 2 8 Date 5/22-5/26/00 Tel Halleris U2723 Concentration FETTIX (entrol D Peri Dish (Mul) No. 04

FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Dala Collected By Steve Turley

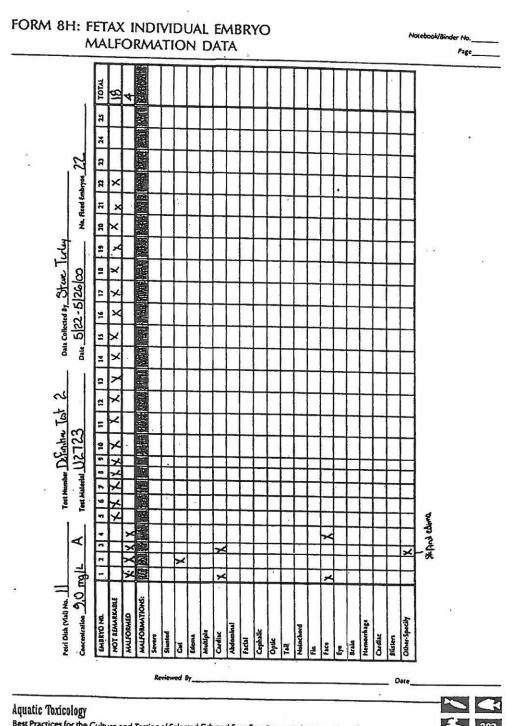


FORM 8H: FETAX INDIVIDUAL EMBRYO Notebook/Binder No. MALFORMATION DATA = Test Malerial U2723 3,2 mg 1L Petri Dlah (Mal) No. 07 Aquatic Toxicology

FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Aquatic Toxicology



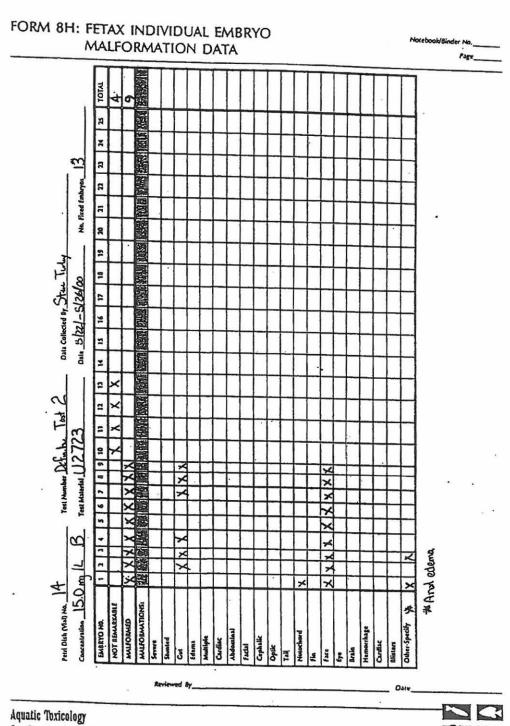
FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA



J

FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Pedi Dish (Mal) No. 9.0 mg/L

FORM 8H: FETAX INDIVIDUAL EMBRYO Notebook/Binder No. MALFORMATION DATA Page Data Collected by Sture Turky 00/42/5-22/5 also Test Muterial 192723 Petri Dish (Mal) Ma. 013



FORM 8H: FETAX INDIVIDUAL EMBRYO Notebook/Binder No. MALFORMATION DATA Page_ col 2/2-2/26/00 25.0 mll

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FORM 8H: FETAX INDIVIDUAL EMBRYO MALFORMATION DATA Date 5/22-5/26/00 Concentration 25.0 my 14

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Hardness

2. 1.6.2		00.1						
Sep. 1893			lour To			9		1
ffluent/Groundwate	r/Toxican	it: Cher	nical (12120			Date: 5	122-5/2
pecies: Xenopus bedi	15			Ac	e at Sta	art of To	ے _aiyst جداد کا عا	ato (Stone
		- 2000 - 020-07		5			These	op Change
Control	Day 0	Day	11	Day	12	Day	y 3	Day 4
FETTAX_	ОН	Old	New	Old	New	Old	New	Old
Temperature (°C)	24.0	24.0	24.0	24.0	24.0	24.0	24.0	24.0
D.O. (mg/L)	8.2	8.3	8.2	7.6	8.1	7.5	8.1	7.6
рН .	-7.14	7.12	7.11	7.18	7.15	7.20.	7.17	7.11
Conductivity								
Alkalinity					4			
Hardness								
		+					9	
19 mg/L	Day 0	T 0:	ay 1	n _a	ıy 2 ·	l Da	ay 3	Day 4
U2723	OH	Old	New	Old	New	Old	New	Old
Temperature (°C)		24.0	590.00000	24.0	240	24.0	240	24.0
D.O. (mg/L)	8.2	8.3		7.8	8.1	7.6	8.2	7.6
рН	7.24	7.17		7.22	7.23	1		7.21
Conductivity	1,01	1.11	1,60	1,00	1.63	1 1.00	1.61	1.61
Alkalinity	1			1		1		†
Hardness	+		,		+	-	1	1
		+	+	+	1-	+	1	
3.2 mg/L	Day 0		Day 1	O	ay 2	0	ay 3	Day 2
U2723	Q.H.	Old	New	Old	· New	- Old	New	Old
Temperature (°C) 24.0	24.0	0 24.0	24.0	24.0	24.0	24.0	24.0
D.O. (mg/L)	8.1	8.0	5.8	7.5	8.0	7.5	1.8	7.5
ρH	7.31	7.2	1 7.28	3 7.23	3 7.30	7.22	7.76	7.7
Conductivity								l.
Alkalinity						1		

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Definitive Test 2
96 Hour Test Data
Page 2 of 3

54 mg/L	Day 0	Day 1		Day 2		Day 3		Day 4	
U2723	ОН	Old	New	Old	New	Old	New	Old	
Temperature (°C)	24.0	24.0	24.0	74.0	24.0	240	24.0	24.0	
D.O. (mg/L)	8.2	8.1	8.2	7.2	8.1	7.6	8.1	7.5	
ρΗ	7.33	7.23	7.35	7.24	7.37	7.25	7.34	7.29	
Conductivity								1	
Alkalinity					Ī				
Hardness		1			İ				
		1			İ			1	
					İ	1	1		

9.0 mg/L	Day 0	Day 1		Day 2		Day 3		Day 4	
U2723	ОН	Old	New	Old	New	Old	New	Old	
Temperature (°C)	24.0	24.0	24.0	24.0	24.0	24.0	24.0	24.0	
D.O. (mg/L)	8.1	8.1	8.1	7.2	8.0	7.7	8.0	7.6	
ρΗ	7.37	7.24	7.35	7.24.	7.38	7.24	7.36	7.33	
Conductivity									
Alkalinity		1							
Hardness									
	1			1	1				
		1				T	1	1.	

15.0 mg/L	Day 0	Day 1		Day 2		Day 3		Day 4	
U2723	OH	Old	New	Old	New	Old	New	Old	
Temperature (°C)	24.0	24.0	24.0	240	24.0	24.0	0.45	24.0	
D.O. (mg/L)	8,2	8.1	8.1	7.2	8.0	7.3	8.1	7.5	
ρΗ	7.40	7.26	7.39	7.23	7.41	7.23	7.40	7.39	
Conductivity									
Alkalinity		1			T				
Hardness	1	1			T				
	1								
•);	1		1						

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DeSintue Test 2

96 Hour Test Data Page 3 of 3

25.0 mg/L	Day 0	Da	Day 1		Day 2		Day 3	
U2723	он	Old	New	Old	New	Old "	New	Old
Temperature (°C)	24.0	240	24.0	24.0	24.0	240	24.0	24.0
D.O. (mg/L)	5.8	8.2	8.2	7.1	8.1	7.2	8.0	7.5
pH .	7.37	7.27	7.34	7.24	7.32	7.25	7.35	7.34
Conductivity	L.							
Alkalinity						190		
Hardness								
	1							

	Day 0	. Day 1		Day 2		Day 3		Day 4
		Old	New	Old	New	Old	New	Old
Temperature (°C)								
D.O. (mg/L)								
рН								
Conductivity					1			
Alkalinity.		-						
Hardness				T				
	1							
						200007100		

	Day 0	Day 1		Day 2		Day 3		Day 4	
		Old	New	Old	New	Old	New	Old .	
Temperature (°C)									
D.O. (mg/L)									
рН									
Conductivity							1		
Alkalinity	(40)	1	1		7				
Hardness		1							
								1	

fluent/Groundwater	Toxican	96 H ا <u>A-</u> ک	our T	est D hand	ata 	(Date:_ <u>5</u>	122-5/26
pecies: Xenopus Im					*	An	alyst: 1	ST
recies. Nortopus Ist	412				je at ot	211 01 11	est:Sby	3.11
intents 5 myle	Day 0	Day	11	Da	y 2	Day	/3	Day 4
(6-AN)	он	Old	New	Old	New	Old	New	Old
Temperature (°C)	24.0	74.0	24.0	24.0	24.0	240	24.0	24.0
0.0. (mg/L)	2.8	8.0	8.1	7.9	8.1	7.8	8.1	7.8
. Ho	-7.10	7.08	7.15	7.11	7.7	7,06	7.13	7.02
Conductivity								
Alkalinity								
Hardness								
	4:							
2500 myle	Day 0	Da	ay 1	Day 2		Day 3		Day 4
(6-An)	он.	Old	New	Old	New	Old	New	Old
Temperature (°C)	74.0	24.0	74.0	24:0	24.0	24:0	24.0	74.0
D.O. (mg/L)	5.8	7.9	8.0		8,0	7.7	8.0	7.6
ρH ·	7.03	7.00	_		1	7,03	7.07	6.98
Conductivity -			1		T			
Alkalinity					T			
Hardness		1						
						1		
			•					
	Day 0	1 7	Day 1	7	Day 2	1	Day 3	Day 4
	0.H	Old		_		_	New	
Temperature (°C		+	-	1	+-	+	+	
D.O. (mg/L)	' - : :	+-	+			+	_	
рН	 	+	+	-		i		—
Conductivity	_	_	+	-	+	1		
Alkalinity	+	+	-		1	+	1	
Hardness	+	+	+	+-	+	1	-	
	+	-	_	- 	-	_	_	
	+	-	-	_	-	1	_	
<u> </u>				23				